



Original Research Article

The protective role of antioxidants extracted from spirulina against hormonal disruption in rabbit ovaries

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Abstract: The study was conducted on 24 adult female local rabbits, which were divided into four groups (6 rabbits per group) and treated as following: the first group received normal saline solution (0.9% sodium chloride) as a control; the second group received valproic acid (400 mg/kg); the third group was treated with spirulina extract only (500 mg/kg); and the fourth group received valproic acid (400 mg/kg) and spirulina extract (500 mg/kg). All treatments were administered for 21 days. Ovarian hormones levels (estrogen and progesterone), as well as anterior pituitary hormones (luteinizing hormone (LH) and follicle-stimulating hormone (FSH)), were assessed using ELISA technique, and histological sections of the ovaries were examined. The results showed a significant decrease in LH, FSH, estrogen, and progesterone levels, and an increase in testosterone levels in the valproate group compared to the control group. Spirulina also had a protective effect against the effects of valproate on the hormone levels under investigation, furthermore, the results showed that spirulina extract reduced ovarian tissue damage caused by valproate. We conclude that spirulina acts as a natural antioxidant, which may reduce reproductive hormonal imbalances and improve damaged ovarian tissue.

Keywords: spirulina, Valproic acid, Reproductive hormones, female rabbits.

Aim of study

to assess if an aqueous extract of *Spirulina* algae can lessen the negative effects on the hormonal and histological characteristics of rabbit ovaries.

INTRODUCTION

Valproic acid (VPA) is an eight-carbon simple branched-chain carboxylic acid synthesized by Burton in 1882, also called 2-propyl pentanoic o (Safdar and Ismail, 2023). It is a broad-spectrum antiepileptic drug and is generally used as monotherapy or in combination with other antiepileptic drugs (Singh *et al.*, 2021).

A series of studies reported that hyperandrogenism,

menstrual disorders, polycystic ovary and other reproductive endocrine abnormalities were more prevalent in women taking VPA (Wood *et al.*, 2005 ; De vries *et al.*,2007).

The study of microalgae has gained importance in recent years, because microalgae are considered as raw materials for chemical compounds that result from their primary and secondary treatment (Chun *et al.*,2011). Microalgae biomass presents remarkable commercial interest in the food, nutraceutical,

cosmetic and pharmaceutical fields due to its high value-added bioactive components (Khan *et al.*, 2018). The discovery of high levels of natural antioxidants in microalgae has triggered the nutraceutical industries to use microalgae-derived metabolites as a replacement for synthetic antioxidants (Azaman *et al.*, 2017).

Antioxidants are substances that limit the effects of free radicals, which are compounds that damage living cells and may cause their death. (Venables *et al.*, 2008; Ahlam & Alaa, 2021). Spirulina is a free-floating filamentous microalgae with the helical properties of its filaments. It is officially called Arthrospira powder and is related to the class of cyanobacteria with the ability to photosynthesize (Sapp, 2005). Spirulina is known for its wide-ranging biological and antioxidant activities (Miranda *et al.*, 1998), so spirulina is considered a valuable source of antioxidants such as phycocyanin, carotenoids, and phenolic compounds (Ismaiel *et al.*, 2016). These biologically active compounds in Spirulina reduce the risk of chronic diseases such as cardiovascular diseases, inflammatory diseases, type 2 diabetes, and cancer (Rodriguez *et al.*, 2018).

This study was designed to determine the effect of spirulina extract on reproductive hormones and related hormones in female rabbits treated with valproate and its effect on ovarian tissue.

MATERIAL AND METHODS

Preparation of the aqueous extract of spirulina:

The cold aqueous extract of spirulina algae was prepared by taking 50 grams of dried algae powder and adding 500 ml of distilled water to it and placing the mixture in a magnetic stirrer for 24 hours, then filtering it using filter paper, then pouring the liquid after filtration into Petri dishes and leaving it to dry at laboratory temperature. Then the algae extract was collected in sterile opaque glass containers and placed in the refrigerator at a temperature of (4) °C until use (Harborn, 1984; Ahmed & Marwa, 2020).

Depakine (valproic acid)

Depakine tablets were powdered, dissolved in distilled water, and administered orally to animals in the form of an emulsion.

Experimental Animals

The study was conducted on female rabbits aged 24 weeks, with an average body weight ranging from 2000 to 2500 g. The rabbits were housed in special cages with specified dimensions. The cages were

cleaned and provided with appropriate environmental conditions, including a fixed lighting regimen (12 h light / 12 h dark), a ventilation cycle, and a temperature ranging from 22 to 25 °C. Adequate amounts of water and feed were supplied in the feeding area inside the cages.

Experimental design:

In the present study, 24 female rabbits were divided into four groups according to the following order:

1. Control group: Consisting of (six) rabbits treated with normal saline NaCl (0.5 ml).
2. Valproate group: Consisting of (six) rabbits given Valproate at a dose of (400 mg/kg) of body weight (Elwakkad *et al.*, 2008).
3. The Third group: Consisting of (six) rabbits treated with *Spirulina* algae extract at a dose of (500mg/kg) of body weight.
4. The Fourth group: consisting of (six) rabbits was Valproate is given as (400mg/kg) of body weight that were treated with *Spirulina* extract (500mg/kg) of body weight.

Blood collection and laboratory tests

After the experimental period, the animals were dissected and blood was collected using a fine needle (5 cc) via cardiac puncture. The collected blood samples were placed in a gel tube for blood collection and allowed to coagulate at room temperature. They were then separated by centrifugation for 15 minutes at 3000 rpm. The serum was then collected and stored in a plain tube at -20°C until use. Hormonal tests were performed using ELISA technology (Aliah *et al.*, 2025), which includes: estrogen, progesterone, luteinizing hormone and follicle-stimulating hormone levels by using (kits of FineTest, China). During the dissection of experimental laboratory animals, we took sections of the ovaries and fixed them with a 10% formalin solution for preservation purposes, and the tissue sections of the ovaries were prepared according to the method (AL-Salmi, 2021).

Statistical analysis:

The results were represented as mean $\pm$ standard deviation ($M \pm SD$), and the experiments were analyzed using one-way analysis of variance (ANOVA) with SPSS and the LSD test, with a p-value < 0.05 .

RESULT

1-Hormonal profile :

1- Follicle-stimulating hormone (FSH)

Figure 1 shows a significant decrease ($p < 0.05$) in follicle-stimulating hormone (FSH) level in the valproic acid group compared to the control group and other groups. However, the spirulina extract (500 mg/kg) group and the valproic

acid + spirulina (500 mg/kg) group showed no significant difference ($p < 0.05$) compared to the control group.

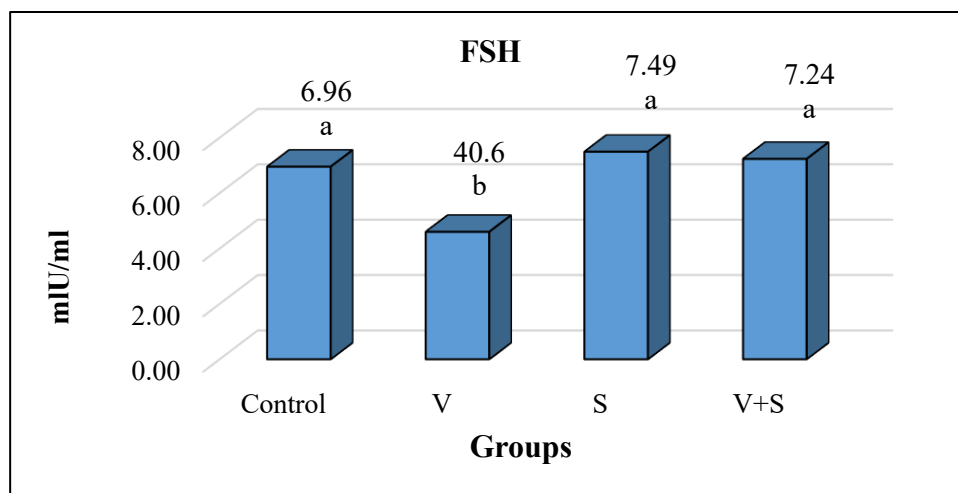


Figure 1: Effect of spirulina extract on FSH level in female rabbits treated with valproic acid.

^{a-b} Different letters indicate significant difference ($P < 0.05$)

2- Luteinizing hormone (LH)

Results showed a significant decrease ($p < 0.05$) in luteinizing hormone (LH) level in the valproic acid group compared to the control group and other groups. However, the valproic acid group combined with spirulina (500 mg/kg) showed no significant difference ($p < 0.05$) compared to the control group. The results also showed a significant increase in LH levels in the spirulina extract group (500 mg/kg) alone (Figure 2).

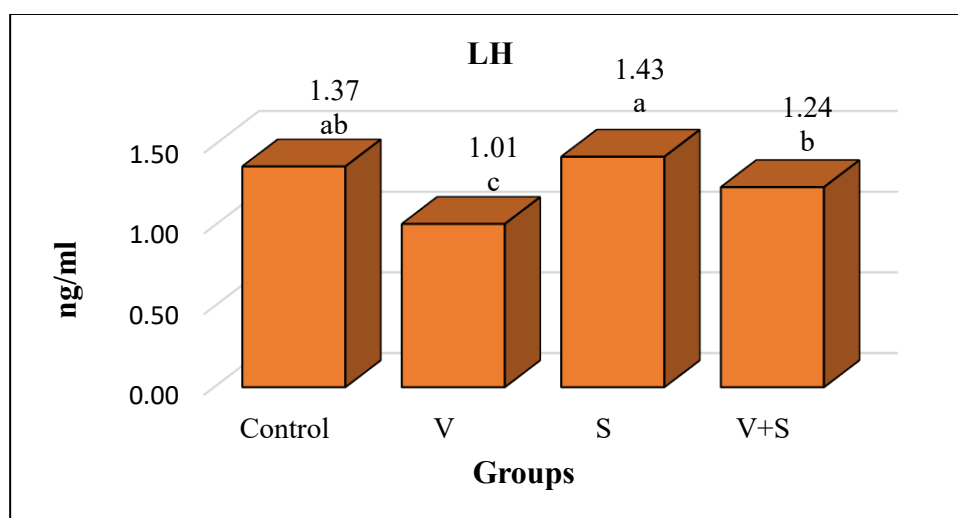


Figure 2: Effect of spirulina extract on LH level in female rabbits treated with valproic acid.

^{a-c} Different letters indicate significant difference ($P < 0.05$)

3- Estrogen hormone (E2)

Figure 3 shows a significant decrease ($p < 0.05$) in estrogen (E2) level in the valproic acid group compared to the control group and other groups. There was a significant increase ($p < 0.05$) in both the spirulina extract (500 mg/kg) group alone and the valproic acid with spirulina (500 mg/kg) group compared to the control group. The greatest increase in hormone levels was observed in the spirulina extract (500 mg/kg) group alone.

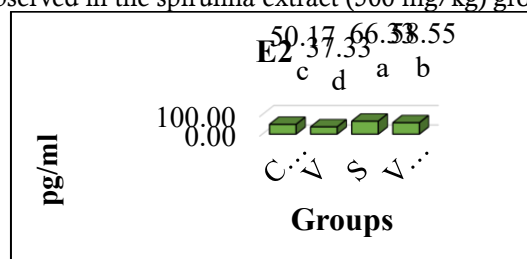


Figure 3: Effect of spirulina extract on E2 level in female rabbits treated with valproic acid.

^{a-d} Different letters indicate significant difference ($P < 0.05$)

4- progesterone hormone

Results showed a significant decrease ($p < 0.05$) in progesterone levels in the valproic acid group compared to the control group and other groups. However, the spirulina extract (500 mg/kg) group and the valproic acid with spirulina (500 mg/kg) group showed no significant difference ($p < 0.05$) compared to the control group (Figure 4).

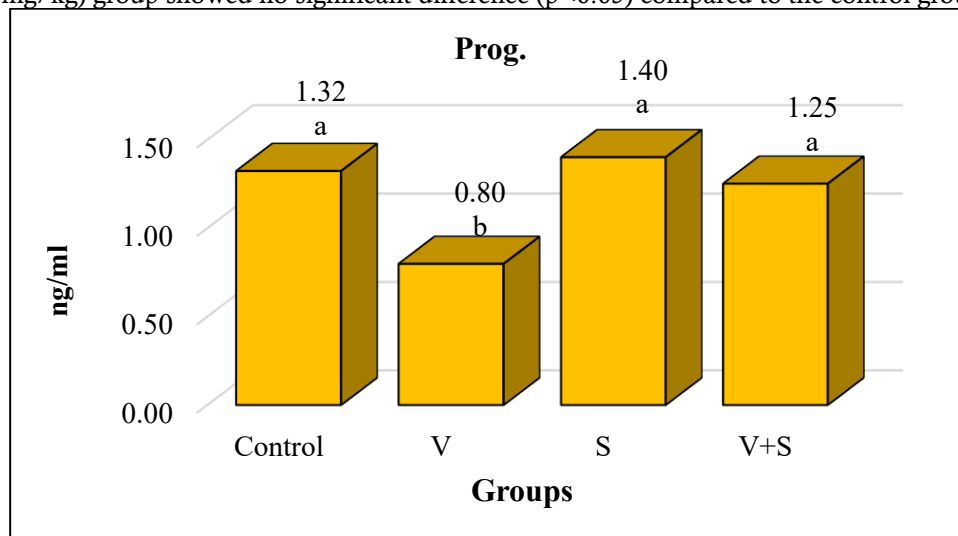


Figure 4: Effect of spirulina extract on Progesterone level in female rabbits treated with valproic acid.

^{a-b} Different letters indicate significant difference ($P < 0.05$)

5- Testosterone hormone

Figure 5 shows a significant increase ($p < 0.05$) in testosterone level in the valproic acid group compared to the control group and other groups. while, the spirulina extract (500 mg/kg) group and the valproic acid with spirulina (500 mg/kg) group showed no significant difference ($p < 0.05$) compared to the control group.

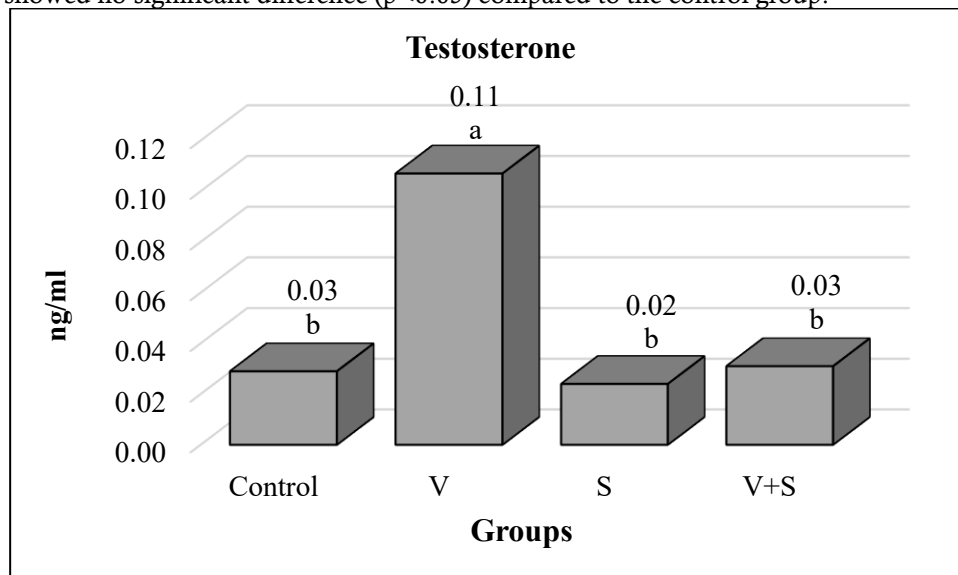


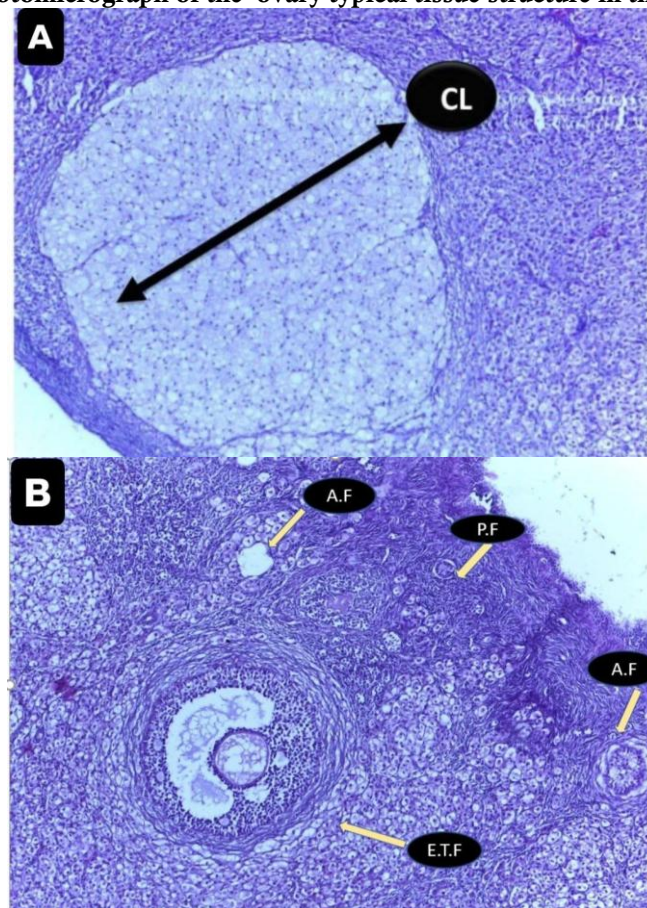
Figure 5: Effect of spirulina extract on Testosterone level in female rabbits treated with valproic acid.

^{a-b} Different letters indicate significant difference ($P < 0.05$)

2-Histological examination:

The histological sections of the ovaries in the control group showed normal follicles at various growth stages, including primordial, primary, secondary, and well-developed graafian follicles corpora lutea were present with normal structure, and the ovarian stroma appeared normal follicles were enclosed with a thin layer of surrounding cells ovaries were covered with a simple cuboidal epithelial tissue graafian follicles were surrounded by mural granulosa cells enclosing the oocyte, with several antral cavities (antra) visible. The sections also displayed stromal fibroblasts and smooth muscle cells. Oocytes were surrounded by a corona radiata, and follicles were embedded within internal and external theca layers, intermixed with fibroblast-like cells the medulla (**Figure 1**).

Figure 1: A photomicrograph of the ovary typical tissue structure in the control group.



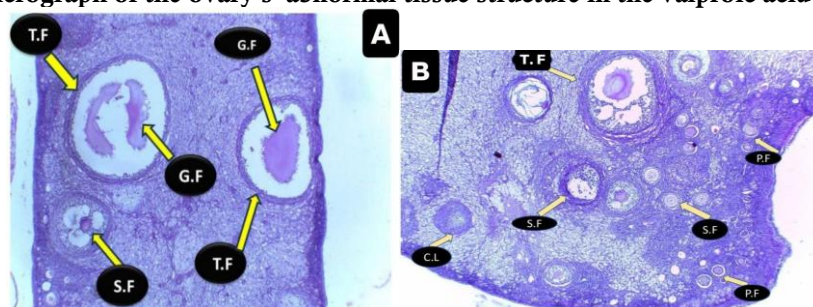
A: showing the Corpus Luteum (C.L). H&E : 100 x

B: Showing the Early Tertiary Follicle (E. T. F) ; Atretic Follicle (A. F) ; Early Atretic Follicle (EA. F) and Primary Follicle (P. F). H&E : 100 x.

The histological examination of rabbit ovaries treated with VPA showed marked degenerative changes. These changes included a decrease in the number of follicles at different developmental stages (primary and secondary follicles), accompanied by an increase in ovarian follicular atresia, additionally, there was evidence of hyperplasia and vascular congestion, along with a reduction in the number of corpora lutea (CL) in the ovarian sections (as seen in figure 2). Degenerated oocytes and atretic follicles were observed, as well as degeneration of granulosa cells showing pyknotic nuclei within the follicular cavity, moreover, connective tissue appeared intermingled with the theca interna, and the zonapellucida was disrupted. The blood vessels were dilated and congested between the stromal cells

The main histological changes in the ovaries of the VPA group indicated a clear impairment of follicular activity, manifested by a small number of developing follicles and a high number of atretic and degenerative follicles, with lysed oocytes

Figure 2: A photomicrograph of the ovary's abnormal tissue structure in the valproic acid group

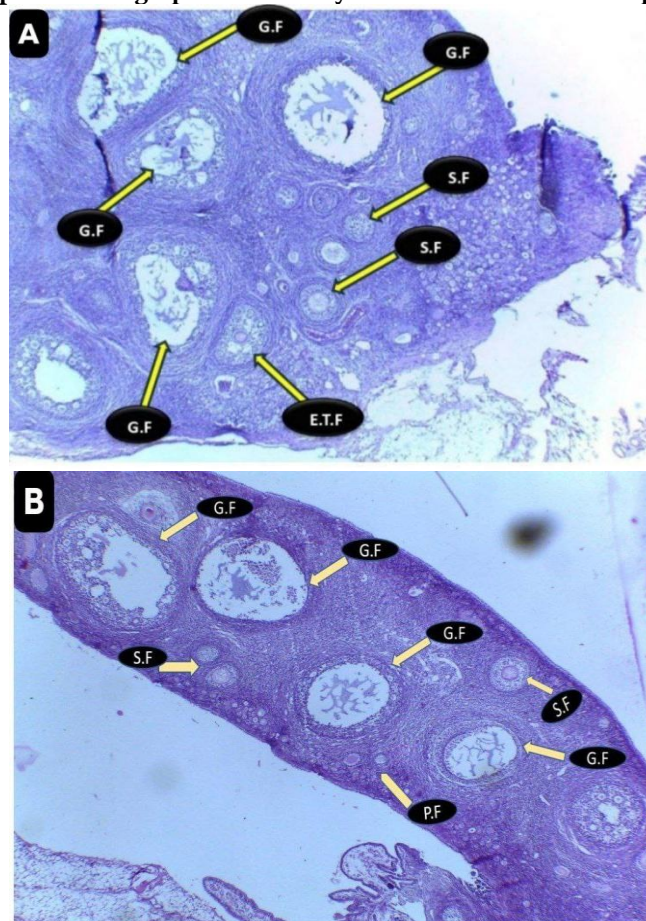


A: Showing the Graafian Follicle (G. F) ; Tertiary Follicle (T. F) and Secondary Follicle (S. F) . H&E : 100 x

B: Showing the Corpus Luteum (C. L) ; Tertiary Follicle (T. F) ; Secondary Follicle (S. F) and Primary Follicle (P. F) . H&E : 100x

The histological sections of the female ovaries treated with Spirulina extract showed a noticeable number of primary and secondary follicles, as well as a few atretic follicles, when compared to the VPA-treated group , generally, the results of our study revealed that there were no significant histological alterations in the ovaries of rabbits administered with Spirulina extracts no vascular congestion was observed, and the histological architecture appeared close to normal, showing well-organized follicles and corpora lutea the morphology of the follicles appeared mature and well-developed (figure 3).

Figure 3: A photomicrograph of the ovary's tissue structure in the spirulinagroup.

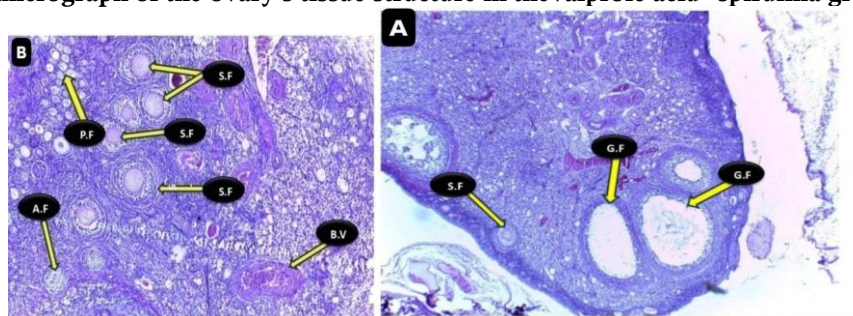


A : Showing the Graafian Follicle (G. F) ; Secondary Follicle (S. F) and Early Tertiary Follicle (E. T. F) . H&E : 100x

B : Showing the Graafian Follicle (G. F) ; Secondary Follicle (S. F) and Primary Follicle (P. F) . H&E : 100x

In summarized description, the section of the female ovaries treated with (Spirulina+ VPA) showed a reduction in congestion of blood vessels, the section also revealed light activity of folliculogenesis, moreover, the ovarian tissue appeared to contain many follicles varying in size and developmental stages, as shown in (figure 4).

Figure 4: A photomicrograph of the ovary's tissue structure in the valproic acid+spirulina group.



A: Showing the Graafian Follicle (G. F) ; Secondary Follicle (S. F) . H&E : 100x

B : Showing the Secondary Follicle (S. F) and Primary Follicle (P. F) ; Atretic Follicle (A. F) and Blood Vessel (B. V) . H&E : 400x

DISCUSSION

The results of this study showed a clear decrease in the levels of FSH and LH in the VAP group compared to the control group. This agrees with (Ibrahim et al., 2019), who reported a significant decrease in FSH and LH levels in female rats treated with VAP.

This potential decrease in hormone levels is attributed to the fact that VPA may lead to an increase in the neurotransmitter gamma-aminobutyric acid (GABA). An increase in GABA can affect the hypothalamus by altering the secretion of gonadotropin-releasing hormone (GnRH), which in turn may affect gonadotropic secretion decreased, FSH and luteinizing hormone levels (Ernst and Goldberg, 2002). In contrast, the spirulina extract group and the spirulina + VAP group showed a noticeable improvement and increase in FSH and LH levels compared to the VAP-treated group. This is consistent with (Hanafi et al., 2025), who reported the supportive effects of Spirulina extract in increasing FSH and LH levels and supporting reproductive function in female rabbits. This is because spirulina is a source of antioxidants and biologically active compounds (Ismail et al., 2016). This extract may contain bioactive compounds or amino acids similar to a general central nervous system stimulatory neurotransmitter that triggers the pulsatile release of GnRH from the hypothalamus, leading to the pulsatile release of pituitary hormones (Garreau and De Rochambeau, 2003). Also, the results showed a clear decrease in its levels of E2 hormone in the VAP group compared to the control group. This decrease is consistent with the study of (Gregoraszczuk et al., 2000), which reported a similar decrease in E2 levels in wistar female rats treated with VAP, and decrease in levels progesterone hormone in the VAP group compared to the control group. This agrees with the study of (Røste et al., 2002), which reported a similar decrease in levels of progesterone in female rats treated with VAP. While regarding testosterone hormone, the results showed a marked increase in the VAP group compared to the control group, this agrees with the study of (Røste et al., 2002), which reported a clear increase in testosterone levels in female rats treated with VAP. Steroid synthesis and ovulation are key functions of the mammalian ovary, and these processes are closely related to follicular formation (Ahn et al., 2021). The inhibitory effect of valproate on cytochrome P450 and glucuronide systems leads to elevated serum testosterone concentrations, the effect of valproate on the ovaries may be due to its effect on the activity of the P450 aromatase enzyme. (Verrotti et al., 2016).

The follicles in women with polycystic ovary syndrome (PCOS) were negative for the P450 aromatase enzyme, thus reducing the conversion of testosterone to estrogen (Tamura et al., 1993).

In contrast, the spirulina extract group and the spirulina + VAP group showed an increase in E2 levels compared to the VAP group. This agrees with the study of (Embaby et al., 2025), and this effect is attributed to the antioxidant compounds present in Spirulina.

Also, the spirulina extract group and the spirulina + VAP group showed a noticeable improvement in progesterone levels compared to the VAP group. This agrees with the study of (Hussin et al., 2022), and this is due to the direct effect of spirulina on hormone secretion from the follicles and its content of essential amino acids, fatty acids, vitamins, and minerals, which is attributed to its antioxidant activity (Hoswini et al., 2013; Farag et al., 2019).

In contrast, the results of the spirulina group and the spirulina + VAP group showed a noticeable improvement in testosterone levels compared to the VAP group. This agrees with the studies of (Bobescu et al., 2020; EL Leithy et al., 2022), which reported that spirulina has an effective role in significantly reducing testosterone levels in female rats, due to the active role of spirulina and its anti-androgenic effect. spirulina is an algae rich in vitamins and minerals, thus exhibiting strong antioxidant activity. It is rich in flavonoids and phenols, which are reflected in its role in improving hormone levels (Hanafi et al., 2025).

Our histological studies of the ovaries confirmed that the improvement in hormone levels due to spirulina showed a moderate effect on ovarian tissue, especially in ovaries treated with valproate, in terms of the growth of mature follicles and corpus luteum, consistent with normal conditions compared to control animals. Perhaps a longer treatment period with spirulina is needed for its positive effect on the ovary tissue to be fully apparent, which is consistent with (Hanafi et al. 2025) who demonstrated the effect of administering an aqueous extract of spirulina on the reproductive capacity of female rabbits; an increase in serum luteinizing hormone and estradiol concentrations was observed.

Conclusion

Research findings indicate that valproic acid exerts significant effects on hormonal balance and ovarian tissue in female rabbits. Notably, the combined administration of valproic acid and Spirulina algae extract resulted in a significant increase in the levels of several reproductive hormones, including luteinizing hormone (LH), follicle-stimulating hormone (FSH), progesterone (P4), and estradiol (E2), accompanied by a marked decrease in testosterone levels. These findings suggest a potential role of spirulina extract in mitigating the hormonal disturbances induced by valproic acid. Furthermore, statistical analyses,

particularly analysis of variance (ANOVA), demonstrated significant differences in hormone levels among the different treatment groups.

References

- Ahn, H. J., An, B. S., Jung, E. M., Yang, H., Choi, K. C., & Jeung, E. B. (2012). Parabens inhibit the early phase of folliculogenesis and steroidogenesis in the ovaries of neonatal rats. *Molecular Reproduction and Development*, 79(9), 626–636.
- Al-Salmi, K. A. (2021). The role of cerium oxide nanoparticles (CeO₂) and β -carotene in normal and cancerous cell lines and laboratory mice: A toxicological, histopathological, and immunohistochemical study (PhD Dissertation). College of Education for Pure Science, University of Basrah, Iraq.
- Azaman, S. N. A., Nagao, N., Yusoff, F. M., Tan, S. W., & Yeap, S. K. (2017). A comparison of the morphological and biochemical characteristics of *Chlorella sorokiniana* and *Chlorella zofingiensis* cultured under photoautotrophic and mixotrophic conditions. *Peer Journal*, 5, 3473–3476.
- Bobescu, E., Bălan, A., Moga, M. A., Teodorescu, A., Mitrică, M., & Dima, L. (2020). Are there any beneficial effects of spirulina supplementation for metabolic syndrome components in postmenopausal women? *Marine Drugs*, 18(12), 651.
- Chun-Yen, C., Kuei-Ling, Y., Rifka, A., Duu-Jong, L., & Jo-Shu, C. (2011). Cultivation, photobioreactor design and harvesting of microalgae for biodiesel production: A critical review. *Bioresource Technology*, 102, 71–81.
- de Vries, L., Karasik, A., Landau, Z., Phillip, M., Kiviti, S., & Goldberg-Stern, H. (2007). Endocrine effects of valproate in adolescent girls with epilepsy. *Epilepsia*, 48(3), 470–477.
- El Leithy, A. A., Al-Karmalawy, A. A., Youssif, O. M., Ebrahim, Y. A., Khalifa, A. S., Elkaed, E. B., & Abo-Zeid, F. S. (2022). Spirulina therapeutic potentiality in polycystic ovarian syndrome management using DHEA-induced rat model. *European Review for Medical & Pharmacological Sciences*, 26(8).
- Elwakkad, A. S., El ElShamy, K. A., & Sibaii, H. (2008). Fish liver oil and propolis as protective natural products against the effect of the anti-epileptic drug valproate on immunological markers of bone formation in rats. *Epilepsy Research*, 80, 47–56.
- Embaby, E. M., Elshopakey, G. E., Megahed, A., Rezk, S., Ateya, A., Eldesoqui, M., & Elghareeb, M. M. (2025). Co-administration of Spirulina and L-carnitine preserves ovarian reserve in a rat model of premature ovarian insufficiency via SIRT1 regulation of oxidative stress, inflammation, and apoptosis. *Scientific Reports*, 15(1), 32527.
- Ernst, C. L., & Goldberg, J. F. (2002). The reproductive safety profile of mood stabilizers, atypical antipsychotics, and broad-spectrum psychotropics. *Journal of Clinical Psychiatry*, 63(Suppl 4), 42–55.
- Farag, M. R., Mahmoud, A., Mohamed, E. A., & Kuldeep, D. (2016). Nutritional and healthical aspects of spirulina (*Arthrospira*) for poultry, animals and human. *International Journal of Pharmacology*, 12, 36–51.
- Garreau, H., & De Rochambeau, H. (2003). La sélection des qualités maternelles pour la croissance du lapereau.
- Gregoraszczyk, E., Wójtowicz, A. K., Taubøll, E., & Ropstad, E. (2000). Valproate-induced alterations in testosterone, estradiol and progesterone secretion from porcine follicular cells isolated from small- and medium-sized ovarian follicles. *Seizure*, 9(7), 480–485.
- Hanafi, E. M., Khalifa, W. H., IA, E., Danial, E. N., Ramadan, M. M., Ezzo, O. H., ... & Mdboli, A. E. N. A. (2025). The role of nano-Spirulina platensis in enhancement of female rabbit puberty and amelioration of the heat stress adverse effects. *Egyptian Journal of Chemistry*, 68(5), 515–526.
- Harborne, J. B. (1984). Methods of plant analysis. In *Phytochemical methods: A guide to modern techniques of plant analysis* (pp. 1–36). Dordrecht: Springer Netherlands.
- Hoseini, S. M., Khosravi-Darani, K., & Mozafari, M. R. (2013). Nutritional and medical applications of spirulina microalgae. *Mini-Reviews in Medicinal Chemistry*, 13(8), 1231–1237.
- Hussin, Z. F., Alwan, N. A., & Abbas, H. K. (2022). Reproductive performance status of adult female rabbits administration Spirulina and combination of folic acid, B6 and B12. *Basrah Journal of Veterinary Research*, 21(S1), 35–43.
- Ibrahim, I. H., Aboregela, A. M., Gouda, R. H. E., & Eid, K. A. (2019). Chronic valproate treatment influences folliculogenesis and reproductive hormones with possible ameliorating role for folic acid in adult albino rats. *Acta Histochemica*, 121(7), 776–783.
- Ismail, M. M. S., El-Ayouty, Y. M., & Piercey-Normore, M. (2016). Role of pH on antioxidants production by Spirulina (*Arthrospira*) platensis. *Brazilian Journal of Microbiology*, 47, 298–304.
- Khan, M. I., Shin, J. H., & Kim, J. D. (2018). The promising future of microalgae: Current status, challenges, and optimization of a sustainable and renewable industry for biofuels, feed, and other products. *Microbial Cell Factories*, 17, 36.
- Miranda, M. S., Cintra, R. G., Barros, S. B., & Mancini Filho, J. (1998). Antioxidant activity of

- the microalga *Spirulina maxima*. *Brazilian Journal of Medical and Biological Research*, 31, 1075–1079.
22. Rodriguez-Concepcion, M., Avalos, J., Bonet, M. L., Boronat, A., Gomez-Gomez, L., Hornero-Mendez, D., Limon, M. C., Meléndez-Martínez, A. J., Olmedilla-Alonso, B., Palou, A., Ribot, J., Rodrigo, M. J., Zacarias, L., & Zhu, C. (2018). A global perspective on carotenoids: Metabolism, biotechnology, and benefits for nutrition and health. *Progress in Lipid Research*, 70, 62–93.
 23. Røste, L. S., Taubøll, E., Isojärvi, J. I., Pakarinen, A. J., Huhtaniemi, I. T., Knip, M., & Gjerstad, L. (2002). Effects of chronic valproate treatment on reproductive endocrine hormones in female and male Wistar rats. *Reproductive Toxicology*, 16(6), 767–773.
 24. Safdar, A., & Ismail, F. (2023). A comprehensive review on pharmacological applications and drug-induced toxicity of valproic acid. *Saudi Pharmaceutical Journal*, 31(2), 265–278.
 25. Sapp, J. (2005). The prokaryote-eukaryote dichotomy: Meanings and mythology. *Microbiology and Molecular Biology Reviews*.
 26. Singh, D., Gupta, S., Verma, I., Morsy, M. A., Nair, A. B., & Ahmed, A. S. F. (2021). Hidden pharmacological activities of valproic acid: A new insight. *Biomedicine & Pharmacotherapy*, 142, 112021.
 27. Tamura, T., Kitawaki, J., Yamamoto, T., Osawa, Y., Kominami, S., Takemori, S., & Okada, H. (1993). Immunohistochemical localization of 17 α -hydroxylase/C17-20 lyase and aromatase cytochrome P-450 in polycystic human ovaries. *Journal of Endocrinology*, 139(3), 503–NP.
 28. Venables, M. C., Hulston, C. J., Cox, H. R., & Jeukendrup, A. E. (2008). Green tea extract ingestion, fat oxidation, and glucose tolerance in healthy humans. *The American Journal of Clinical Nutrition*, 87(3), 778–784.
 29. Verrotti, A., Mencaroni, E., Cofini, M., Castagnino, M., Leo, A., Russo, E., & Belcastro, V. (2016). Valproic acid metabolism and its consequences on sexual functions. *Current Drug Metabolism*, 17(6), 573–581.
 30. Wood, J. R., Nelson-Degrave, V. L., Jansen, E., McAllister, J. M., Mosselman, S., & Strauss, J. F., 3rd. (2005). Valproate-induced alterations in human theca cell gene expression: Clues to the association between valproate use and metabolic side effects. *Physiological Genomics*, 20(3), 233–243.