



Original Researcher Article

In Vitro Antifungal Efficacy of Acetone Leaf Extracts From Selected Aromatic Plants Against Phytopathogenic Fungi Associated With *Carica Papaya* L.

Article History:

Name of Author:

Chandrakant D. Ghorband¹, Madhuri M. Maindargikar¹, Anil C. Shinde², Rahul V. Zade³, Vaijanath V. Kharat⁴, Gangadhar M. Bhosale⁵, Saheb L. Shinde^{1*}

Affiliation: ¹Department of Botany, Yeshwant Mahavidyalaya, Nanded, Dist. Nanded (MH), India.

²Department of Botany, Shri Datta Arts, Commerce and Science College, Hadgaon, Dist. Nanded, (MH), India.

³Department of Botany, Shankarrao Chavan Mahavidyalaya, Ardhapur, Dist. Nanded, (MH), India.

⁴Department of Biology, Yeshwant Mahavidyalaya, Nanded, Dist. Nanded (MH), India.

⁵Department of Botany, Mahatma Gandhi Mahavidyalaya, Ahmedpur, Dist. Latur, (MH), India.

Corresponding Author: Saheb L. Shinde

Received: 02/11/2025

Revised: 30/11/2025

Accepted: 20/12/2025

Published: 28/12/2025

Abstracts: Papaya (*Carica papaya* L.) cultivation is severely affected by fungal diseases, resulting in significant yield losses and increased reliance on chemical fungicides. However, the extensive use of synthetic fungicides raises serious environmental and health concerns highlighting the need for safer and sustainable alternatives. The present study investigated the phytochemical composition and in vitro antifungal efficacy of acetone leaf extracts of five aromatic medicinal plants, namely *Pongamia pinnata* (L.) Pierre, *Melia azedarach* (L.), *Vitex negundo* (L.), *Ocimum sanctum* (L.) and *Tagetes erecta* (L.). Preliminary phytochemical screening confirmed the presence of diverse bioactive compounds, including alkaloids, flavonoids, glycosides, steroids, terpenoids, phenols and tannins, with noticeable variation among species. Antifungal activity was evaluated using the agar well diffusion method against major papaya pathogens, viz., *Fusarium solani*, *Oidium caricae*, *Mycosphaerella caricae*, *Phytophthora palmivora* and *Pythium aphanidermatum*. Among the tested extracts, *Vitex negundo* exhibited the highest inhibitory activity, producing zones of inhibition ranging from 15.33 ± 0.88 to 20.00 ± 0.57 mm, followed closely by *Melia azedarach* 15.66 ± 1.45 to 19.66 ± 0.57 mm. Moderate antifungal effects were observed for *Tagetes erecta*, *Pongamia pinnata* and *Ocimum sanctum*. Although the standard fungicide Bavistin showed superior inhibition, the activity of *V. negundo* and *M. azedarach* was comparable against key pathogens. Overall, the strong antifungal efficacy associated with rich phytochemical content highlights these medicinal plants as promising eco-friendly alternatives to synthetic fungicides for sustainable papaya disease management.

This is an open access journal, and articles are distributed under the terms of the Creative Commons Attribution-Noncommercial-Share Alike 4.0 License, which allows others to remix, tweak, and build upon the work non-commercially, as long as appropriate credit is given and the new creations are licensed under the identical terms.

Keywords: *Carica papaya* L., Phytochemicals, Antifungal activity, Aromatic plants, sustainable agriculture.

INTRODUCTION

Carica papaya L. (papaya) is a highly valued tropical fruit crop cultivated extensively in India and other tropical and subtropical regions due to its rapid growth, high productivity and exceptional nutritional and medicinal value. India ranks among the leading global producers of papaya contributing substantially to domestic consumption and international trade. Papaya fruits are rich sources of essential vitamins (A, C, and E), minerals, dietary fiber and bioactive compounds, including carotenoids, phenolics and digestive enzymes such as papain which confer antioxidant, anti-inflammatory, digestive and immunomodulatory properties (Saran et al., 2014; Fuentes and Santamaria et al., 2014; Sanikommu, et al., 2021). Owing to these attributes papaya occupies a prominent position in nutritional security, traditional medicine, and agro-based economies.

Despite its economic and dietary significance, papaya production is severely constrained by fungal diseases that cause considerable pre- and post-harvest losses, leading to deterioration in fruit quality, reduced shelf life and significant economic setbacks for farmers. Major phytopathogenic fungi associated with papaya include *Fusarium solani*, *Oidium caricae*, *Mycosphaerella caricae*, *Phytophthora palmivora* and *Pythium aphanidermatum* which are responsible for destructive diseases such as root rot, damping-off, powdery mildew, fruit rot, leaf spot and stem canker (Rawal, 2010; Srikantharajah, et al., 2021; Guang Heng, et al., 2023; Getnet, et al., 2024). These infections may result in yield losses ranging from 30% to 70%, depending on climatic conditions, cultivar susceptibility, and disease severity (Fenta, et al., 2023; Peja et al., 2025). The intensification of fungal epidemics under changing climatic patterns has further exacerbated the need for effective sustainable disease management strategies.

Conventional disease control in papaya largely relies on chemical fungicides which, although effective in the short term, pose serious concerns including environmental pollution, accumulation

of toxic residues in fruits, disruption of beneficial soil microflora and the emergence of fungicide-resistant pathogen strains (Chavez-Quintal, et al., 2011; Ademe, 2013; Atul et al., 2021; Deresa, and Diriba, 2023). These drawbacks have intensified global efforts to develop eco-friendly, biodegradable and plant-based alternatives for crop protection. Botanical extracts derived from medicinal and aromatic plants have emerged as promising natural fungicides due to their broad-spectrum antimicrobial activity, environmental safety and minimal risk to human health (Zaker, 2016; G Sandhya, et al., 2024).

Medicinal plants such as *Pongamia pinnata*, *Melia azedarach*, *Vitex negundo*, *Ocimum sanctum* and *Tagetes erecta* are well recognized in traditional Indian medicine and agriculture for their antimicrobial, insecticidal and therapeutic properties. Their antifungal efficacy is attributed to diverse phytochemicals, including alkaloids, flavonoids, phenols, tannins, terpenoids and coumarins which disrupt fungal cell membrane integrity, inhibit spore germination and interfere with metabolic and enzymatic processes (Tiwari and Prajapati, 2024; Pattanayak, et al., 2010; Vaibhav et al., 2024; Madhavi and Revathi, 2025; Singh, et al., 2020). Recent studies have demonstrated significant antifungal activity of plant extracts against major phytopathogens, including *Fusarium*, *Colletotrichum*, *Phytophthora* and *Pythium* species underscoring their potential in sustainable agriculture (Honnesh and Saha, 2021; Fugare, et al., 2021; Deresa and Diriba, 2023).

Plants are rich natural sources of bioactive compounds with proven antimicrobial potential and long-standing use in traditional medicine. Growing concerns over environmental safety and fungicide resistance have accelerated the search for eco-friendly alternatives to synthetic pesticides. Fungal diseases cause major yield and economic losses in papaya cultivation, highlighting the urgent need for sustainable disease management strategies. Plant-derived phytochemicals offer a promising, biodegradable, and safer approach for controlling

phytopathogenic fungi and improving crop protection (Oniha, et al., 2021; Ayilara, et al., 2023; Santra et al., 2020).

Although numerous plant species have been individually screened for antifungal properties comparative studies focusing on acetone leaf extracts of selected aromatic plants against major papaya phytopathogens remain limited. Acetone is considered an efficient extraction solvent capable of dissolving a wide spectrum of polar and non-polar phytochemicals, thereby enhancing antifungal efficacy (Ademe, 2013; Dissanayake, et al., 2019; Sharmila, et al., 2020). Therefore, the present study was undertaken to evaluate the in vitro antifungal activity and phytochemical composition of acetone leaf extracts of *Pongamia pinnata*, *Melia azedarach*, *Vitex negundo*, *Ocimum sanctum* and *Tagetes erecta* against key fungal pathogens associated with *Carica papaya* L. The findings aim to contribute toward the development of eco-friendly, sustainable and economically viable plant-based fungicides for integrated disease management in papaya cultivation.

MATERIALS AND METHODS

Collection of plant materials

The leaves of five selected aromatic plant species namely Karanj (*Pongamia pinnata* L.) Pierre, Nimbora (*Melia azedarach* L.), Nirgudi (*Vitex negundo* L.), Tulsi (*Ocimum sanctum* L.), and Zendu (*Tagetes erecta* L.), were collected from the rural regions of Kandhar, District Nanded, Maharashtra, India, during their active growing season. Only fresh and healthy leaves were selected to ensure maximum phytochemical integrity. The collected plant materials were carefully identified and authenticated in the department of botany Yeshwant Mahavidyalaya, Nanded (Kokate, 2010; Harborne, 1998). After collection, the leaves were thoroughly washed under running tap water followed by rinsing with distilled water to eliminate dust and surface impurities. The cleaned samples were then shade-dried at ambient room temperature for a period of 7–10 days until complete dehydration was achieved. The dried leaves were finely powdered using a sterile mechanical grinder to obtain a uniform particle size. The powdered samples were transferred into airtight containers and stored at 4 °C to prevent degradation and preserve their bioactive constituents until further experimental use (Sasidharan et al., 2011).

Preparation of plant extracts

50 g of powdered plant materials was extracted using 250 mL of acetone in a Soxhlet extraction

apparatus for 6–8 hours with the temperature maintained at 56°C (boiling point of acetone). (Harborne, 1998; Azwanida, 2015). The extract was filtered through Whatman No. 1 filter paper and the solvent was evaporated under reduced pressure using a rotary evaporator to obtain crude extracts. The dried extracts were weighed and stored in sterile glass vials at 4 °C. Stock solutions (100 mg/mL) were prepared using dimethyl sulfoxide (DMSO) and sterilized through a 0.22 µm membrane filter and concentrated using appropriate method (Nostro et al., 2000).

Isolation and identification of fungal pathogens

Diseased papaya plant samples (leaves, fruits, and stems) showing typical fungal infection symptoms were collected from papaya fields. The samples were surface sterilized using 1% sodium hypochlorite for 2–3 minutes, followed by triple washing with sterile distilled water. Small tissue segments were aseptically inoculated onto Potato Dextrose Agar (PDA) medium and incubated at 27 ± 2 °C for 5–7 days (Dhingra and Sinclair, 1995). Pure cultures were obtained by subculturing actively growing hyphal tips and identified based on colony morphology and microscopic features using standard mycological manuals (Barnett and Hunter, 1998; Leslie and Summerell, 2006). The fungal species identified included *Fusarium solani*, *Oidium caricae*, *Mycosphaerella caricae*, *Phytophthora palmivora*, and *Pythium aphanidermatum*.

Antifungal activity assay

The antifungal activity of acetone leaf extracts was evaluated using the agar well diffusion method (Perez et al., 1990; Valgas et al., 2007). Sterile molten PDA medium was poured into Petri plates and allowed to solidify. A standardized fungal spore suspension (10⁶ spores/mL) was uniformly spread on the surface of the medium. Wells of 6 mm diameter were aseptically punched, and 100 µL of plant extract at different concentrations (25, 50, 75, and 100 mg/mL) was added into each well. DMSO served as the negative control, while Bavistin (0.1%) was used as the standard control. Plates were incubated at 27 ± 2 °C for 72–96 hours. The antifungal activity was assessed by measuring the diameter of inhibition zones (mm). All experiments were performed in triplicate.

Phytochemical analysis

Preliminary phytochemical analysis of acetone leaf extracts was performed to detect the presence of glycosides, alkaloids, flavonoids, steroids, terpenoids, saponins, coumarin, proteins, reducing sugar, phenols and tannins using standard

qualitative procedures (Harborne, 1998; Kokate, 2010; Sofowora, 2008). The analysis included tests for glycosides using Salkowski Test, alkaloids using Mayer's test, Flavonoids using Shinoda test, Steroids using Chloroform test, Terpenoids using Salkowaski test, Saponins using Foam test, Coumarin using Coumarin test, Proteins using Biuret test, Reducing Sugar using Fehling test, Phenols and Tannins using Ferric Chloride test.

Statistical Analysis

All experiments were conducted in triplicate, and data were expressed as mean $\pm$ standard deviation (SD). Statistical significance was determined using one-way analysis of variance (ANOVA) followed by Tukey's post hoc test at $p < 0.05$ using SPSS software (Gomez and Gomez, 1984; Steel et al., 1997).

RESULT AND DISCUSSION

Phytochemical analysis of Selected aromatic plant extracts

Table 1: Phytochemical constituents of acetone leaf extracts of selected aromatic plants

Phytochemical Constituents	Test	<i>Pongamia pinnata</i> (L.) Pierre	<i>Melia azedarach</i> (L.)	<i>Vitex negundo</i> (L.)	<i>Ocimum sanctum</i> (L.)	<i>Tagetes erecta</i> (L.)
Glycosides	Salkowski Test	++	+	+	++	++
Alkaloids	Mayer's test	+++	++	++	+++	++
Flavonoids	Shinoda test	+++	+++	++	++	+++
Steroids	Chloroform test	+++	++	+	+	+
Terpenoids	Salkowaski test	++	++	++	+	++
Saponins	Foam test	+	+	+	+	+
Coumarin	Coumarin test	++	++	+	+	++
Proteins	Biuret test	+	+	+	++	+
Reducing Sugar	Fehling test	+	+	+	++	+
Phenols	Ferric Chloride test	++	++	++	+++	+++
Tannins	Ferric Chloride test	++	+++	+++	+++	+++

Where: - + In low concentration, ++ In moderate concentration, +++ In high concentration and - Absent,

The preliminary phytochemical screening of acetone leaf extracts from the selected aromatic plants *Pongamia pinnata*, *Melia azedarach*, *Vitex*

negundo, *Ocimum sanctum* and *Tagetes erecta* revealed the presence of a wide range of bioactive constituents, although their distribution and relative abundance varied among species in Table:1).

Glycosides were detected in all five plant extracts with *Pongamia pinnata*, *Ocimum sanctum* and *Tagetes erecta* exhibiting moderate levels while *Melia azedarach* and *Vitex negundo* showed comparatively lower concentrations. Alkaloids were abundantly present in *Pongamia pinnata* and *Ocimum sanctum* whereas moderate levels were observed in *Melia azedarach*, *Vitex negundo* and *Tagetes erecta*. Flavonoids were prominently detected in *Pongamia pinnata*, *Melia azedarach* and *Tagetes erecta*, indicating their rich antioxidant potential, while moderate amounts were recorded in *Vitex negundo* and *Ocimum sanctum*.

Steroids were found in high concentration in *Pongamia pinnata* moderate levels in *Melia azedarach* and low levels in the remaining plant species. Terpenoids were moderately present in *Pongamia pinnata*, *Melia azedarach*, *Vitex negundo* and *Tagetes erecta* whereas *Ocimum sanctum* exhibited only a low concentration. Saponins were detected in low amounts across all the studied plant extracts suggesting their limited yet consistent presence.

Coumarins were moderately present in *Pongamia pinnata*, *Melia azedarach* and *Tagetes erecta* while *Vitex negundo* and *Ocimum sanctum* showed comparatively lower levels. Proteins and reducing sugars were detected in low to moderate concentrations with *Ocimum sanctum* showing relatively higher levels than the other plant species. Phenolic compounds were abundant in *Ocimum sanctum* and *Tagetes erecta* whereas moderate levels were observed in the remaining species. Similarly, tannins were found in high concentrations in *Melia azedarach*, *Vitex negundo*, *Ocimum sanctum* and *Tagetes erecta*, while *Pongamia pinnata* showed moderate content.

The present investigation revealed a rich and diverse phytochemical profile in acetone leaf extracts of *Pongamia pinnata*, *Melia azedarach*, *Vitex negundo*, *Ocimum sanctum* and *Tagetes erecta* with

glycosides, alkaloids, flavonoids, steroids, terpenoids, phenols and tannins consistently detected. The relative abundance of these bioactive components among plant species appears to significantly influence their antifungal potential. In particular, *Vitex negundo* and *Melia azedarach* the high levels of phenols, flavonoids, alkaloids, and tannins found in *Melia azedarach* are strongly associated with its excellent antifungal efficacy which supports previous reports linking the potent antifungal and antibiofilm activities of this species to its phenolic-rich composition (Sharmila et al., 2020; Lakshmisha et al., 2020; Jaiswal and Kumar, 2025; Akacha et al., 2022). Similarly, the increased polyphenolic content in *T. erecta* reinforces its recognized antifungal and biopesticidal potential, as documented in recent studies showing inhibition of the growth of phytopathogenic fungi (Birson et al., 2022; Martinez-Ruiz et al., 2024). Although *Pongamia pinnata* and *Ocimum sanctum* exhibited relatively moderate antifungal activity, its rich content of flavonoids, phenols, tannins, and alkaloids underscores its therapeutic importance, corroborating previous findings on its antimicrobial properties and traditional medicinal relevance (Sharmila et al., 2020; Verma and Pandey, 2023). Taken together, the present findings confirm that the synergistic action of multiple secondary metabolites plays a significant role in antifungal efficacy and highlight the potential of this aromatic medicinal plant as an eco-friendly alternative to synthetic fungicides for sustainable plant disease management.

Overall, the rich diversity of phytochemical constituents observed in these plants supports their traditional medicinal use and suggests their strong potential as natural sources of antimicrobial, antioxidant and biopesticidal agents. The presence of key secondary metabolites such as alkaloids, flavonoids, phenols, tannins and terpenoids may be responsible for the observed biological activities of these plant extracts.

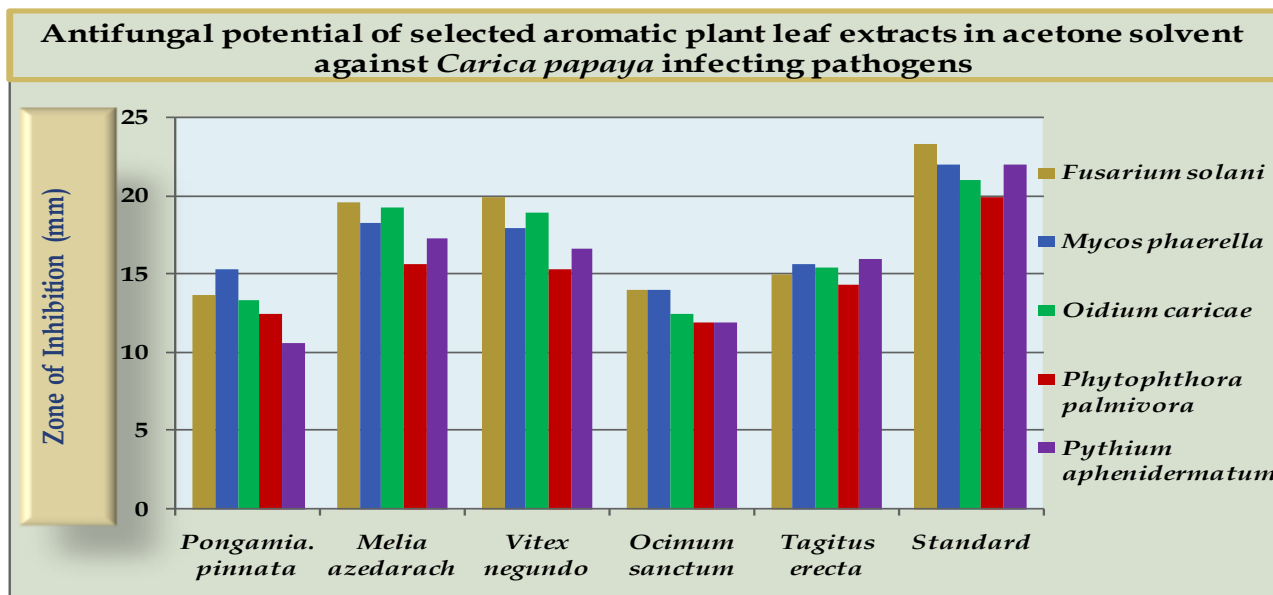
In vitro antifungal activity of selected aromatic plant extracts

Table 2: Antifungal potential of selected aromatic plant leaf extracts in acetone solvent against Papaya (*Carica papaya* L.) infecting pathogens.

Sr. No	Selected Medicinal Plants	Zone of Inhibitions in mm				
		<i>Fusarium solani</i>	<i>Oidium caricae</i>	<i>Mycosphaerella caricae</i>	<i>Phytophthora palmivora</i>	<i>Pythium aphenidermatum</i>
1	<i>Pongamia pinnata</i>	13.66±1.20	15.33±1.33	13.33±0.72	12.50±1.04	10.66±0.92
2	<i>Melia azedarach</i>	19.66±0.57	18.33±0.57	19.33±1.33	15.66±1.45	17.33±0.16
3	<i>Vitex negundo</i>	20.00±0.57	18.00±1.04	19.00±1.15	15.33±0.88	16.66±0.16
4	<i>Ocimum sanctum</i>	14.00±1.15	14.00±1.15	12.50±0.50	12.00±0.76	12.00±1.15
5	<i>Tagetis erecta</i>	15.00±1.15	15.66±0.66	15.50±1.04	14.33±0.57	16.00±0.76
6	Standard (Bavistin)	23.33±0.72	22.00±0.76	21.00±0.57	20.00±0.76	22.00±0.57

Note: Values are mean ± SD (n = 3). Different superscripts in a column indicate significant differences (p < 0.05).

Figure 1: Antifungal potential of selected aromatic plant leaf extracts in acetone solvent against Papaya (*Carica papaya* L.) infecting pathogens.



The antifungal activity of five aromatic plant leaf extracts prepared using acetone solvent, namely *Pongamia pinnata* (L.), Pierre, *Melia azedarach* (L.), *Vitex negundo* (L.), *Ocimum sanctum* (L.), *Tagetis erecta* (L.) and tested against five selected common fungal pathogens *Fusarium solani*, *Mycosphaerella caricae*, *Oidium caricae*, *Phytophthora palmivora* and *Pythium aphenidermatum* of *Carica papaya* plant and their fruits using the agar well diffusion assay.

Bavistin had been used as a control (fungicide) to compare the effect of plant based treatments with synthetic fungicides. The data on the effect of aromatic plant extracts on plant pathogenic fungi of *Carica papaya* plant and their fruits are presented in Table 2.

The extracts of five aromatic medicinal plant leaves showed significant reduction in the growth of

Fusarium solani, *Mycosphaerella caricae*, *Oidium caricae*, *Phytophthora palmivora* and *Pythium aphanidermatum* and there was significant difference in the efficacy of these extracts. Among the acetone extracts of all five plants, two plants showed greater inhibition of mycelial growth of all pathogenic fungi tested than the control, *Vitex negundo* and *Melia azedarach* showed exceptionally prominent activity. The leaf extract of *Vitex negundo* showed maximum activity 20.00 ± 0.57 mm zone of inhibition against *Fusarium solani* even at low concentrations and *Melia azedarach* also showed 19.66 ± 0.57 mm zone of inhibition. The following plants, such as *Tagetes erecta*, *Pongamia pinnata* and *Ocimum sanctum* exhibited fungicidal activity against all pathogenic fungi tested. Therefore, this study suggests that acetone extracts of the screened plants would be useful for treating plant and fruit diseases of *Carica papaya* caused by *Fusarium solani*, *Mycosphaerella caricae*, *Oidium caricae*, *Phytophthora palmivora* and *Pythium aphanidermatum*. The Bavistin used as a standard antifungal drug as control showed inhibition against the tested fungi, it also showed antifungal activity at 5 µg concentration.

Similar antifungal trends against phytopathogens associated with papaya using plant-based extracts have been studied by Patil et al., (2021), who reported strong inhibition of *Fusarium* and *Phytophthora* species under Indian agro-climatic conditions. Similarly, Karthikeyan et al., (2022) shows that acetone and methanol extracts of *Vitex negundo* and *Tagetes erecta* significantly suppressed fungal pathogens of tropical fruit crops, resulting in

CONCLUSION

The findings of this study clearly indicate that acetone leaf extracts of selected aromatic medicinal plants contain a broad spectrum of phytochemicals and exhibit strong antifungal activity against major fungal pathogens of papaya. The consistent detection of biologically active secondary metabolites, particularly alkaloids, flavonoids, phenols, tannins and terpenoids, appears to play a central role in mediating the fungicidal effects. Among the species evaluated, *Vitex negundo* and *Melia azedarach* exhibited excellent inhibitory potential, inducing fungicidal responses, underscoring their promise as effective natural fungicidal agents. The remarkable fungicidal efficacy of these plant extracts not only supports their traditional medicinal relevance but also emphasizes their practical value in developing sustainable plant disease management strategies. The use of plant-based formulations can

high phenolic and flavonoid contents. Jadhav et al., (2023) confirmed the efficacy of *Melia azedarach* extracts against soil and fruit-borne fungi, highlighting their role in sustainable disease management. Additionally, Deshmukh et al., (2020) reported remarkable antifungal activities of *Ocimum sanctum* and *Pongamia pinnata* against *Fusarium oxysporum* and *Pythium aphanidermatum*, supporting the broad-spectrum antifungal potential of these plants. Similarly, Pawar and Patil, (2022) observed significant inhibition of the growth of papaya phytopathogens using botanical extracts of medicinal plants highlighting their utility in biological disease control strategies. More recently, Chavan et al., (2024) reported that phytochemical-rich leaf extracts of indigenous plants effectively suppressed fungal pathogens of horticultural crops, further reinforcing the importance of botanical fungicides in reducing the dependence on chemical fungicides. Taken together, these findings strongly corroborate current results and underscore the growing scientific consensus that plant-based antifungal agents are environmentally friendly, biodegradable, and safe alternatives to synthetic fungicides for the sustainable management of papaya diseases.

It is clear that the acetone extract of leaves of *Vitex negundo* and *Melia azedarach* and other tested plants should be used as a potent biopesticide to treat plant diseases of *Carica papaya* plant and their fruits caused by fungi as it shows maximum activity even at almost the same low concentrations as standard antifungal agents.

significantly reduce the dependence on chemical fungicides, thereby reducing environmental pollution reducing the level of toxic residues in agricultural products, and reducing the risks of pathogen resistance development. Consistent with previous studies highlighting the effectiveness of botanical extracts and local biocontrol agents in crop protection, the current study further strengthens the scientific basis for the use of plant-derived antifungal agents as environmentally safe, biodegradable and economically viable alternatives. Overall, this study provides a strong scientific framework for advancing ecofriendly botanical fungicides and makes a significant contribution to the search for sustainable, safe and residue-free papaya production systems.

ACKNOWLEDGEMENT

Authors would like to thank Principal Yeshwant Mahavidyalaya, Nanded for providing research facilities.

REFERENCES

- Ademe, Anteneh. (2013). Evaluation of Antifungal Activity of Plant Extracts against Papaya Anthracnose (*Colletotrichum gloeosporioides*). *Journal of Plant Pathology & Microbiology*, 04, 10.
- Akacha, N., Rebiai, A., Lanez, T., & Djebbar, M. R. (2022). Antifungal activity of ethanolic extracts of *Melia azedarach* leaves against phytopathogenic fungi. *Journal of Plant Protection Research*, 62(2), 156–164.
- Atul, Kumar & Singh, Jai & Singh, Pooja & Javeria, Shaily. (2021). Postharvest Diseases of Papaya and Their Management. 10.1201/9781003045502-20.
- Ayilara, M. S., Adeleke, B. S., Akinola, S. A., Fayose, C. A., Adeyemi, U. T., Gbadegesin, L. A., Omole, R. K., Johnson, R. M., Uthman, Q. O., & Babalola, O. O. (2023). Biopesticides as a promising alternative to synthetic pesticides: A case for microbial pesticides, phytopesticides, and nanobiopesticides. *Frontiers in microbiology*, 14, 1040901.
- Azwanida, N. N. (2015). A review on the extraction methods use in medicinal plants, principle, strength and limitation. *Medicinal & Aromatic Plants*, 4(3), 196.
- Barnett, H. L., & Hunter, B. B. (1998). *Illustrated genera of imperfect fungi* (4th ed.). APS Press.
- Bîrsan, C., Muntean, D., Marian, E., & Tofană, M. (2022). Phytochemical characterization and antifungal potential of *Tagetes erecta* extracts against major plant pathogens. *Plants*, 11(4), 478.
- Chavan, A. M., Jadhav, S. A., & Patil, D. S. (2024). Evaluation of indigenous medicinal plant extracts against fungal pathogens of horticultural crops. *Indian Phytopathology*, 77(1), 112–120.
- Chávez-Quintal, P., González-Flores, T., Rodríguez-Buenfil, I., & Gallegos-Tintoré, S. (2011). Antifungal Activity in Ethanolic Extracts of *Carica papaya* L. cv. Maradol Leaves and Seeds. *Indian journal of microbiology*, 51(1), 54–60.
- Deresá, E. M., & Diriba, T. F. (2023). Phytochemicals as alternative fungicides for controlling plant diseases: A comprehensive review of their efficacy, commercial representatives, advantages, challenges for adoption, and possible solutions. *Heliyon*, 9(3), e13810.
- Deresá, E. M., & Diriba, T. F. (2023). Phytochemicals as alternative fungicides for controlling plant diseases: A comprehensive review of their efficacy, commercial representatives, advantages, challenges for adoption, and possible solutions. *Heliyon*, 9(3), e13810.
- Deshmukh, A. J., Pawar, P. V., & Patil, S. R. (2020). Antifungal activity of selected medicinal plant extracts against important phytopathogenic fungi. *Journal of Pharmacognosy and Phytochemistry*, 9(4), 1896–1901.
- Dhingra, O. D., & Sinclair, J. B. (1995). *Basic plant pathology methods* (2nd ed.). CRC Press.
- Fenta, L., Mekonnen, H., & Kabtamer, N. (2023). The Exploitation of Microbial Antagonists against Postharvest Plant Pathogens. *Microorganisms*, 11(4), 1044.
- Fuentes, Gabriela & Santamaría, Jorge. (2014). *Papaya (Carica papaya L.): Origin, Domestication, and Production*. 10.1007/978-1-4614-8087-7_1.
- Fugare, A. G., Shete, R. V., Adak, V. S., & G., K. M. (2021). A Review on *Pongamia pinnata* (L.): Traditional Uses, Phytochemistry and Pharmacological Properties. *Journal of Drug Delivery and Therapeutics*, 11(1-s), 207-211.
- G Sandhya, Divya Shree, DH Tandel, Vinita Yadav and G Punith (2024). Studies on biochemical changes and eco-friendly management of post-harvest anthracnose in papaya. *Int. J. Adv. Biochem. Res.*8(8):1263-1268.
- Getnet, Muluken & Alemu, Kumlachew & Tsedale, Binyam. (2024). Status of postharvest papaya anthracnose (*Colletotrichum gloeosporioides*) in Assosa Zone, Western Ethiopia. *Discover Food*. 4.
- Gomez, K. A., & Gomez, A. A. (1984). *Statistical procedures for agricultural research* (2nd ed.). John Wiley & Sons.
- Grover, R. K., & Moore, J. D. (1962). Toximetric studies of fungicides against brown rot organisms, *Sclerotinia fructicola* and *S. laxa*. *Phytopathology*, 52, 876–880.
- Guang Heng, Tan & Ali, Asgar & Siddiqui, Yasmeen. (2023). Major fungal postharvest diseases of papaya: Current and prospective diagnosis methods. *Crop Protection*. 174.
- Harborne, J. B. (1998). *Phytochemical methods: A guide to modern techniques of plant analysis* (3rd ed.). Chapman & Hall.
- Jadhav, S. A., Patil, D. S., & Chavan, A. M. (2023). Bioefficacy of selected botanicals

- against fungal pathogens of fruit crops. *Indian Journal of Agricultural Research*, 57(4), 498–504.
24. Jaiswal, S., & Kumar, V. (2025). Phenolic profiling and antifungal potential of *Vitex negundo* L. against *Candida albicans*. *Journal of Ethnopharmacology*, 325, 116921.
 25. Karthikeyan, V., Ganesan, S., & Kumar, R. S. (2022). Evaluation of antifungal potential of medicinal plant extracts against major plant pathogens. *Indian Journal of Natural Products and Resources*, 13(2), 210–218.
 26. Kokate, C. K. (2010). *Practical pharmacognosy* (4th ed.). Vallabh Prakashan.
 27. Lakshmeesha, T. R., Kalagatur, N. K., Mudili, V., & Lee, Y. R. (2020). Antifungal and antiaflatoxigenic activities of *Melia azedarach* leaf extracts and silver nanoparticles. *Journal of Applied Microbiology*, 128(4), 1214–1226.
 28. Lakshmeesha, T. R., Sateesh, M. K., Daruka Prasad, B. K., & Chandrasekhar, S. (2020). Antifungal activity of *Melia azedarach* leaf extract and its silver nanoparticles against *Fusarium* species. *Journal of Applied Microbiology*, 128(2), 543–556.
 29. Leslie, J. F., & Summerell, B. A. (2006). *The Fusarium laboratory manual*. Blackwell Publishing.
 30. M, Madhavi & Koyya, Revathi. (2025). Evaluation of antibacterial and antifungal activity of *Ocimum sanctum* leaf extract. *Journal of Innovations in Applied Pharmaceutical Science (JIAPS)*.
 31. Maha Laksha Mudiyanseleage Chandrika Dissanayake, Villani Sepala Dahanayaka and Chithrani Ranasinghe, 2019. Antifungal Potential of Some Plant Extracts Against *Colletotrichum gloeosporioides* Causal Organism of Papaya Anthracnose Disease. *Asian Journal of Biological Sciences*, 12: 589-595.
 32. Martínez-Ruiz, N. R., Hernández-Carranza, P., López-Hernández, L. H., & Espinosa-Andrews, H. (2024). Polyphenol composition and biological activity of *Tagetes erecta* extracts. *Industrial Crops and Products*, 198, 116774.
 33. N. H., Honnesh, and Santanu Saha. 2021. "Antibacterial and Antifungal Activity of Selected Indian Traditional Medicinal Plants". *Journal of Pharmaceutical Research International* 33 (60B):623-34.
 34. Nostro, A., Germano, M. P., D'Angelo, V., Marino, A., & Cannatelli, M. A. (2000). Extraction methods and bioautography for evaluation of medicinal plant antimicrobial activity. *Letters in Applied Microbiology*, 30(5), 379–384.
 35. Oniha, M., Eni, A., Akinola, O., Omonigbehin, E. A., Ahuekwe, E. F., & Olorunshola, J. F. (2021). In vitro Antifungal Activity of Extracts of *Moringa oleifera* on Phytopathogenic Fungi Affecting *Carica papaya*. *Open Access Macedonian Journal of Medical Sciences*, 9(A), 1081–1085.
 36. Patil, S. R., Pawar, P. V., & Jadhav, R. S. (2021). Antifungal activity of selected plant extracts against phytopathogens of papaya. *Indian Phytopathology*, 74(3), 645–652.
 37. Pattanayak, P., Behera, P., Das, D., & Panda, S. K. (2010). *Ocimum sanctum* Linn. A reservoir plant for therapeutic applications: An overview. *Pharmacognosy reviews*, 4(7), 95–105.
 38. Pawar, R. D., & Patil, S. R. (2022). Botanical extracts as eco-friendly fungicides for management of papaya diseases. *Indian Journal of Agricultural Research*, 56(3), 356–361.
 39. Peja Jr, R., Dimayacyac, D. A., Barcelo, J., De Torres, R., Mandap, J. A., Amas, J., ... & Balendres, M. A. O. (2025). Fungi in the Postharvest of Agricultural Commodities. In *Mycology in a Changing Planet: Applications and Perspectives from Southeast Asia* (pp. 245-300).
 40. Perez, C., Pauli, M., & Bazerque, P. (1990). An antibiotic assay by the agar well diffusion method. *Acta Biologica et Medicinæ Experimentalis*, 15, 113–115.
 41. Rawal, R. D. (2010). Fungal diseases of papaya and their management. *Acta Horticulturae*, 851, 443–446.
 42. Sanikommu, Vijay Rakesh Reddy & Aj, Sachin & Chinnasamy, Kavitha & Kalal, Prashant. (2021). *Papaya (Carica papaya L.)*.
 43. Santra, H. K., & Banerjee, D. (2020). Natural Products as Fungicide and Their Role in Crop Protection. *Natural Bioactive Products in Sustainable Agriculture*, 131–219.
 44. Saran, Parmeshwar Lal & Choudhary, Ravish & Solanki, Ishwar & Kumar, P R. (2014). New fruit and seed disorders in Papaya (*Carica papaya* L.) in India. *African Journal of Biotechnology*. 13. 574-580.
 45. Sasidharan, S., Chen, Y., Saravanan, D., Sundram, K. M., & Yoga Latha, L. (2011). Extraction, isolation and characterization of bioactive compounds from plants' extracts. *African Journal of Traditional*,

- Complementary and Alternative Medicines, 8(1), 1–10.
46. Sharmila, D., E. Kowsalya, and R. Kamalambigeswari. 2020. “Antifungal Activity of Medicinal Plant Extracts Against Plant Pathogen *Colletotrichum* Sp”. *Plant Cell Biotechnology and Molecular Biology* 20 (23-24):1341-44.
 47. Sharmila, G., Muthukumaran, C., Suresh Kumar, N., & Thirumarimurugan, M. (2020). Antifungal activity and phytochemical screening of selected medicinal plants against phytopathogens. *Journal of Applied Biology & Biotechnology*, 8(4), 44–52.
 48. Singh, J., & Tripathi, N. N. (1999). Inhibition of storage fungi of blackgram (*Vigna mungo*) by some essential oils. *Flavour and Fragrance Journal*, 14(1), 1–4.
 49. Singh, Yogendra & Gupta, Amit & Kannoja, Pushpendra. (2020). *Tagetes erecta* (Marigold) - A review on its phytochemical and medicinal properties. *Current Medical and Drug Research*. 4. 10.53517/CMDR.2581-5008.412020201.
 50. Sofowora, A. (2008). *Medicinal plants and traditional medicine in Africa* (3rd ed.). Spectrum Books Limited.
 51. Srikantharajah, N., Pakeerathan, K., & Mikunthan, G. (2021). Fungicide-Free Management of Papaya Anthracnose (*Colletotrichum gloeosporioides* Penz.) Disease Using Combined Bio-Rationales and Bee Wax in Organic Agriculture. *Biology and Life Sciences Forum*, 4(1), 16.
 52. Steel, R. G. D., Torrie, J. H., & Dickey, D. A. (1997). *Principles and procedures of statistics: A biometrical approach* (3rd ed.). McGraw-Hill.
 53. Tiwari, G. C., & Prajapati, M. (2024). *Melia azedarach* L.: A plant with a wide range of phytopharmacological properties. *African Journal of Biomedical Research*, 27(6S), 843–848.
 54. Vaibhav Paymal*, Jaya Kambale, Dr. Nilesh Chougule, Antifungal Property of Medicinal Plants of *Vitex Negundo*, *Acacia Nilotica* and *Hibiscus Rosa Sinesin*, *Int. J. of Pharm. Sci.*, 2024, Vol 2, Issue 11, 1578-1589.
 55. Verma, S., & Pandey, A. K. (2023). Phytochemical profiling and antimicrobial evaluation of *Pongamia pinnata* leaf extracts. *South African Journal of Botany*, 155, 410–419.
 56. Verma, S., & Pandey, R. (2023). Antimicrobial potential of *Pongamia pinnata* and its phytochemical constituents. *Journal of Pharmacognosy and Phytochemistry*, 12(1), 146–152.
 57. Zaker, M. (2016). Natural Plant Products as Eco-friendly Fungicides for Plant Diseases Control- A Review. *The Agriculturists*, 14(1), 134–141.