



Original Research Article

Cytokines as a therapeutic target for rheumatoid arthritis

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Abstract:

Rheumatoid arthritis (RA) is a chronic autoimmune inflammatory condition characterized by synovial membrane inflammation, leading to joint damage and functional impairment. Cytokines play a pivotal role in the pathogenesis of RA, orchestrating a complex network of immune cells and inflammatory processes. This review article explores the efficacy and safety of targeting cytokines as a therapeutic approach for RA, focusing on major cytokines including tumor necrosis factor-alpha (TNF- α), interleukin-6 (IL-6), interleukin-1 (IL-1), interleukin-17 (IL-17), interleukin-23 (IL-23), granulocyte macrophage-colony stimulating factor (GM-CSF), and other cytokines investigated as potential targets. For this review, literature was systematically searched using databases such as PubMed and Scopus, ensuring a comprehensive overview of existing studies. This article aims to provide unique insights into the current landscape of cytokine-targeted therapies for RA, distinguishing it from other reviews by not only summarizing clinical outcomes but also critiquing the complexities of cytokine interactions and the lessons learned from failed therapeutic targets. The review discusses preclinical rationale, clinical outcomes of biologic agents targeting TNF- α , IL-6, IL-1, IL-17, IL-23, GM-CSF, and side effects of these therapies, along with the challenges in translating preclinical findings into clinical benefits.

In conclusion, targeting cytokines remains a promising and evolving strategy in the management of RA, offering potential avenues for improving patient outcomes and quality of life while emphasizing the need for personalized approaches in treatment.

Keywords: Rheumatoid arthritis, cytokines, TNF- α , IL-6, IL-1, IL-17, IL-23, GM-CSF, autoimmune disease, biologic therapy, inflammation, targeted therapy

INTRODUCTION

Rheumatoid arthritis (RA) is characterized as an autoimmune disorder that primarily affects the synovial joints, leading to systemic inflammation and joint destruction. The autoimmune nature of RA indicates that an inappropriate immune response targets the body's own tissues, particularly the joints. Evidence suggests that genetic factors, such as specific alleles of the major histocompatibility complex (MHC), in conjunction with environmental factors (e.g., infections, tobacco use), can trigger this immune response, resulting in the production of autoantibodies and the activation of autoreactive T and B cells [1,2].

The signs and symptoms of RA are typically the result of synovitis—an inflammation of the synovial membrane of the joints. This inflammatory process involves a complex network of cells and cytokines, particularly in the recruitment, activation, and effector functions of immune cells [3,4]. In addition to the influx of B and T lymphocytes, plasma cells, mast cells, dendritic cells, and neutrophils, fibroblasts also infiltrate the synovial membrane. The hypertrophied synovial layer, known as pannus, contributes to the destruction of bone and cartilage in the joint. Interleukin-1 (IL-1) was the first cytokine detected in the synovial fluid of patients with RA and has been associated with cartilage degradation in vitro [5,6].

Subsequently, levels of tumor necrosis factor-alpha (TNF-α) were also shown to correlate with cartilage degradation in vitro. TNF-α is a potent pro-inflammatory cytokine that induces several other cytokines in the pro-inflammatory cascade, including IL-1, IL-6, IL-8, and granulocyte macrophage-colony stimulating factor (GM-CSF), and increases the levels of various adhesion molecules such as intracellular adhesion molecule (ICAM) and vascular cell adhesion molecule (VCAM) [7,8]. Structural erosions in RA

occur at the border between the pannus (the hypertrophied layer of synovial cells) and cartilage, in an area densely populated by TNF-α-producing cells. It has been shown that the neutralization of TNF-α using anti-TNF-α antibodies in mouse models of arthritis results in a significant reduction in joint edema and structural erosions [9].

In RA, pro-inflammatory cytokines released by activated immune cells play a crucial role in mediating the autoimmune response, perpetuating inflammation, and contributing to destructive processes within the joints. This review will focus on elucidating the mechanisms by which these cytokines contribute to RA pathogenesis, as well as the potential side effects of cytokine inhibitors [10,11].

CYTOKINE TARGETS

Cytokines have long been explored as potential targets for rheumatoid arthritis (RA) because they are directly involved in the disease process, which can be classified into pro-inflammatory and anti-inflammatory cytokines based on their functions in response to antigens. A seminal preclinical study demonstrated the utility of blocking TNF-α in RA, showing that antibodies against TNF-α significantly reduced high IL-1 production in RA synovial cell cultures [12,13].

Pro-inflammatory cytokines such as TNF-α, IL-1β, IL-6, IL-7, IL-15, IL-17, IL-18, IL-23, IFN-γ, and GM-CSF play critical roles in regulating inflammation and are present at elevated levels in the synovium, synovial fluid, serum, or peripheral blood of RA patients. Certain cytokines, including IL-15, IL-17, IL-23, and GM-CSF, are closely linked to rheumatoid factor (RF), anti-cyclic citrullinated peptide (CCP) seropositivity, and RA activity, making them potential diagnostic biomarkers. Notably, IL-7 is recognized for its role in early RA, with levels varying across different disease stages [14-16].

Table 1. Key Cytokines Involved in Rheumatoid Arthritis and Their Targets

Cytokine	Role in RA	Therapeutic Target	Example Inhibitor	Key Effects
TNF-α	Promotes inflammation and joint destruction	TNF-α receptor inhibition	Adalimumab	Reduces inflammation, joint swelling, and pain
IL-6	Mediates systemic inflammation	IL-6 receptor inhibition	Tocilizumab	Decreases CRP levels, improves physical function
IL-1	Induces synovial inflammation	IL-1 receptor inhibition	Anakinra	Reduces inflammatory response, slows joint damage
IL-17	Enhances inflammation and joint destruction	IL-17 receptor inhibition	Secukinumab	Reduces disease activity
IL-23	Supports Th17 cell maintenance and IL-17 production	IL-23 receptor inhibition	Ustekinumab	Improves clinical symptoms, reduces inflammation
GM-CSF	Stimulates myeloid cell activity	GM-CSF receptor inhibition	Mavrilimumab	Decreases disease activity, pain, disability
IL-10	Anti-inflammatory cytokine	IL-10 receptor/agonist targeting	Dekavil	Enhances regulatory T cells

Additionally, macrophages secrete various cytokines, including TNF- α , which promotes the proliferation of fibroblast-like synoviocytes (FLS) and synovial cells through the activation of nuclear factor kappa-light-chain-enhancer of activated B cells (NF- κ B) and ERK-1/2-E26 pathways. This activation leads to the production of inflammatory mediators such as IL-6, matrix metalloproteinase-1 (MMP-1), and MMP-3. Here, the redundancy in cytokine roles becomes apparent; IL-1 β enhances MMP production and leukocyte adhesion in RA joints by activating ERK, JNK, AP-1, and NF- κ B, while IL-6 induces bone resorption and cartilage degradation via MMPs and receptor activator of nuclear factor kappa-B ligand (RANKL) [17]. IL-7 and IL-15 are implicated in T-cell trafficking and antigen-specific CD4⁺ T-cell proliferation, respectively, with IL-7 showing potential as a biomarker for early RA. Furthermore, IL-18 and IL-12 jointly stimulate interferon-gamma (IFN- γ) production, further promoting inflammation [18].

Targeting these cytokines with inhibitors has proven effective in reducing symptoms and halting disease progression. For example, adalimumab, a TNF- α inhibitor, prevents TNF- α from binding to its receptors, thereby reducing inflammation and protecting cartilage and bone from destruction. However, the potential side effects of these therapies—including increased susceptibility to infections, malignancies, and infusion reactions—necessitate careful monitoring and consideration in treatment plans to ensure patient safety [19].

Conversely, anti-inflammatory cytokines such as IL-4, IL-10, IL-13, and TGF- β counteract the inflammatory process. Understanding these protective roles is essential for their utilization in targeted therapies. For instance, IL-10, produced by regulatory T cells, suppresses Th17 cells and promotes Treg cell development. Exploring the therapeutic potential of these cytokines could enhance existing treatments and underscore the benefits of personalized medicine strategies.

Introducing IL-4, IL-10, IL-13, or TGF- β as therapeutic agents could harness their anti-inflammatory properties to mitigate the symptoms of rheumatoid arthritis (RA). IL-4 exhibits an anti-angiogenic effect through the inhibition of vascular endothelial growth factor (VEGF), while IL-13 provides cartilage protection via downregulation of Fc gamma receptor I (Fc γ RI), further supporting their therapeutic potential. This intricate interplay of cytokines underscores the complexity of RA pathogenesis and the potential of targeted cytokine therapies to improve patient outcomes.

IL-13, a cytokine associated with Th2 cell-mediated immune responses, exerts its anti-angiogenic function by activating protein kinase C (PKC) α/β II and ERK-1/2, while simultaneously downregulating the NF-

κ B/p65 pathway. It may also reduce chondrocyte death and protect cartilage from destruction by lowering Fc γ RI levels [20,21]. Transforming growth factor (TGF)- β , primarily expressed by macrophages and T lymphocytes, promotes the migration and invasion of fibroblast-like synoviocytes (FLS), inducing epithelial-mesenchymal transition (EMT) via Smad-2/3 activation in RA [22]. Clinical trials have shown that dekalil (an IL-10 agonist) demonstrates significant efficacy in RA patients, as these agents can bind to and initiate receptors, inducing corresponding biological responses.

Thus, cytokines play a fundamental role in the pathogenesis of RA through their regulation of inflammation and interactions with various immune cells. Elevated levels of pro-inflammatory cytokines, such as TNF- α , IL-1 β , IL-6, IL-7, IL-15, IL-17, IL-18, IL-23, IFN- γ , and GM-CSF, are found in the synovium, synovial fluid, serum, and peripheral blood of RA patients, highlighting their central role in disease progression [23]. TNF- α , produced by macrophages, stimulates FLS and synovial cells via NF- κ B and Erk-1/2-E26 signaling pathways, leading to the secretion of inflammatory mediators like IL-6, MMP-1, and MMP-3, thereby exacerbating inflammation [11].

The treatment of RA has evolved significantly over the past 20 years. Traditional therapies included glucocorticoids and nonsteroidal anti-inflammatory drugs (NSAIDs) for acute inflammation, alongside disease-modifying antirheumatic drugs (DMARDs) such as methotrexate (MTX), hydroxychloroquine, and sulfasalazine for maintenance therapy [24,25]. DMARDs, particularly MTX, have been shown to reduce joint symptoms and extra-articular manifestations while inhibiting radiographic progression, so they continue to be used as first-line therapies. However, although some patients with RA respond to DMARDs alone, a significant proportion do not tolerate this therapy and/or continue to have active disease despite treatment. Additionally, cumulative side effects, particularly from corticosteroids and NSAIDs, can exacerbate comorbidities. This situation created a need for alternative therapies, leading to the emergence of biologic agents for treating moderate to severe RA in the early 1990s [26,27].

Inhibition of inflammatory cytokines, particularly TNF- α , became a major focus of RA clinical research in the 1990s, a trend that continues today. The first open trial using a TNF- α -blocking drug was conducted in the UK in 1992, where 20 patients with RA received infliximab, a chimeric antibody specific to TNF- α . This treatment resulted in a significant reduction in the signs and symptoms of RA, accompanied by a decrease in inflammatory markers [28]. The early promise of anti-TNF- α therapy in RA has been confirmed in several large multicenter trials, revolutionizing the treatment of RA and providing crucial insights into the disease's

pathophysiology. These therapies have achieved blockbuster status, generating substantial profits for the pharmaceutical industry. Since the advent of infliximab, other anti-TNF- α agents and cytokine inhibitors have been developed and introduced into clinical practice. Although primarily developed and tested in RA, these drugs have also been used in other chronic inflammatory diseases, sometimes yielding unexpected and divergent results [29,30].

ANTI-TUMOR NECROSIS FACTOR

Currently, five TNF- α inhibitors are licensed for the treatment of rheumatoid arthritis (RA). These include three full-size monoclonal antibodies (infliximab, adalimumab, and golimumab), one humanized antigen-binding fragment conjugated with polyethylene glycol (certolizumab), and one soluble fusion protein (etanercept). These biologics target and neutralize tumor necrosis factor-alpha (TNF- α), a key cytokine in the inflammatory processes associated with RA.

The American College of Rheumatology (ACR) has established a core set of seven disease activity indicators: swollen joint count, tender joint count, physician's global assessment of disease activity, patient's global assessment of disease activity, patient's assessment of pain, patient's assessment of physical function, and an acute-phase reactant level (either C-reactive protein [CRP] or erythrocyte sedimentation rate [ESR]) [31,32]. These indicators are used to assess treatment efficacy and were instrumental in developing the ACR response criteria, which are standardized measures of clinical improvement. The ACR20 response—indicating a 20% improvement in tender and swollen joint counts along with three of the other five core measures—serves as the standard primary endpoint in RA clinical trials [33]. ACR20 evaluates individual patient improvement rather than average group improvement, typically reported as the percentage of participants achieving this level of improvement. As treatment modalities have advanced, higher thresholds such as ACR50 and ACR70, reflecting 50% and 70% improvement respectively, have also been incorporated into clinical evaluations [34].

Phase III trials for TNF- α inhibitors, including those approved for RA, consistently demonstrated the primary endpoint (ACR20) in patients with active RA who inadequately responded to methotrexate (MTX-IR) [35]. While these studies showed variable times to achieve the primary endpoint, resulting in higher placebo responses in shorter trials, the overall response rates were similar across different TNF- α inhibitors. In MTX-IR patients, approximately 60%, 40%, and 20% achieved ACR20, ACR50, and ACR70 responses, respectively, by week 24. Despite these promising results, significant unmet needs remain, as about 40% of patients do not achieve even a 20% response, and only a small minority reach remission in the short term. A meta-analysis of 6-month ACR response rates from

16 studies comparing various biologics to a placebo-MTX combination estimated the number needed to treat (NNT) to achieve ACR20, ACR50, and ACR70 responses at 3.2, 4.2, and 7.7, respectively. Long-term studies and systematic reviews have further validated the efficacy of TNF- α inhibitors in reducing radiographic progression of erosive damage and improving quality of life, as measured by the Health Assessment Questionnaire (HAQ) [36].

In clinical practice, TNF- α inhibitors like adalimumab function by binding to TNF- α , thereby preventing it from interacting with its receptors and reducing the inflammatory processes associated with cytokines such as MMP-1 and MMP-3. This inhibition helps prevent cartilage and bone destruction. Despite their effectiveness, a significant portion of RA patients do not respond adequately to TNF- α inhibitors, highlighting the need for personalized treatment approaches and ongoing research into alternative therapeutic targets [37]. The development of additional cytokine inhibitors and biologics targeting different aspects of the immune response is crucial for addressing the diverse and complex nature of RA. Furthermore, the role of anti-inflammatory cytokines such as IL-4, IL-10, IL-13, and TGF- β in counteracting RA inflammation presents potential therapeutic avenues. Introducing these cytokines or their agonists could complement existing therapies, providing a multifaceted approach to RA management and improving patient outcomes [38].

ANTI-INTERLEUKIN-6 (IL-6) DRUGS AND INTERLEUKIN-1 (IL-1) INHIBITORS EFFECTS AND BIOCHEMISTRY

Anti-Interleukin-6 (IL-6) drugs and interleukin-1 (IL-1) inhibitors are crucial for the treatment of rheumatoid arthritis (RA) due to their roles in modulating the immune response and inflammation. IL-6 is a multifunctional cytokine involved in immune regulation, hematopoiesis, and inflammation. It exerts its effects by binding to its receptor, IL-6R, which exists in both membrane-bound and soluble forms [39]. The IL-6/IL-6R complex then associates with the signal-transducing component gp130, triggering downstream signaling pathways, including the JAK-STAT pathway. This activation leads to the transcription of genes involved in inflammatory responses. In RA, IL-6 contributes to synovitis and systemic inflammation by promoting the differentiation of B cells into antibody-producing plasma cells, enhancing T cell activation, and stimulating the production of acute-phase reactants like C-reactive protein (CRP). Elevated IL-6 levels correlate with disease activity and joint destruction [40,41].

Tocilizumab, a humanized monoclonal antibody against the IL-6 receptor, inhibits IL-6 signaling by preventing IL-6 from binding to both soluble and membrane-bound IL-6 receptors. This blockade reduces inflammation, decreases CRP levels, and

improves clinical symptoms in RA patients [42]. Tocilizumab has demonstrated efficacy in patients who inadequately respond to traditional disease-modifying antirheumatic drugs (DMARDs) and TNF- α inhibitors, leading to reduced joint damage progression and improved physical function. Another IL-6 inhibitor, sarilumab, also targets the IL-6 receptor and exhibits similar therapeutic effects [43].

Interleukin-1 (IL-1) is a pro-inflammatory cytokine primarily produced by activated macrophages. It exists in two forms: IL-1 α and IL-1 β , both of which bind to the IL-1 receptor type I (IL-1RI), initiating signal transduction that activates NF- κ B and MAPK pathways. These pathways induce the expression of various inflammatory mediators, including additional cytokines, chemokines, and matrix metalloproteinases (MMPs), contributing to synovial inflammation and cartilage degradation in RA. IL-1 also enhances the differentiation of osteoclasts, leading to bone resorption [44].

Anakinra, a recombinant human IL-1 receptor antagonist, competitively inhibits the binding of IL-1 to IL-1RI, thus blocking IL-1-mediated signaling. This inhibition reduces the inflammatory response and slows the progression of joint damage [45,46]. Although effective, anakinra is less commonly used than other biologics due to its daily injection requirement and more modest efficacy compared to alternative options. Additionally, canakinumab, an IL-1 β monoclonal antibody, and rilonacept, a soluble decoy receptor for IL-1, provide alternative approaches to IL-1 inhibition. While effective in reducing inflammation and disease activity in RA, their use is more limited compared to other biologics [47].

Both anti-IL-6 drugs and IL-1 inhibitors significantly impact RA by modulating key cytokine pathways involved in the disease's pathogenesis. By targeting these cytokines, these therapies help control systemic and local inflammation, reduce joint damage, and improve patient outcomes [48].

TARGETING THE INTERLEUKIN-17/23 CYTOKINE PATHWAY

Targeting the IL-17 and IL-23 cytokine pathway in rheumatoid arthritis (RA) represents an emerging therapeutic strategy due to the significant roles these cytokines play in the disease's inflammatory processes. IL-17, particularly IL-17A, is produced by T helper 17 (Th17) cells and is known to promote inflammation by inducing the production of several pro-inflammatory mediators, including TNF- α , IL-1, and IL-6, as well as chemokines and matrix metalloproteinases (MMPs) [49, 50]. These factors contribute to joint inflammation and destruction.

IL-23, composed of p19 and p40 subunits, is crucial for the differentiation and maintenance of Th17 cells, thereby influencing the production of IL-17 [51]. Clinically, agents that inhibit either IL-17 or IL-23 are being explored for their potential to reduce

inflammation and disease progression in RA. Monoclonal antibodies such as secukinumab and ixekizumab specifically bind to IL-17A, preventing it from interacting with its receptor, thereby blocking its pro-inflammatory effects. Clinical trials indicate that these agents can reduce disease activity and improve clinical symptoms in RA patients [52, 53]. However, the response rates observed in RA have been less robust than those seen in other IL-17-mediated conditions, such as psoriasis and ankylosing spondylitis. This suggests that while IL-17 is involved in RA, its role may not be as dominant as in those conditions or that a more comprehensive blockade of the IL-23/Th17 axis might be necessary [54, 55].

Additionally, ustekinumab, which targets the shared p40 subunit of IL-12 and IL-23, indirectly affects the IL-23/Th17 pathway by inhibiting Th17 cell differentiation and IL-17 production. Clinical trials of ustekinumab in RA have shown some efficacy, resulting in improvements in clinical symptoms and reductions in inflammatory markers, although these results have been less pronounced compared to those seen in other autoimmune diseases [56, 57].

Therapeutic methods targeting the IL-17/23 pathway typically involve the subcutaneous administration of these biologic agents at intervals ranging from two to four weeks, depending on the specific drug and patient response. Such therapies are generally considered for patients who have not adequately responded to traditional treatments or TNF- α inhibitors. While the safety profiles of IL-17 and IL-23 inhibitors are generally favorable, ongoing monitoring for potential side effects, such as an increased risk of infections due to immune modulation, is essential [58, 59].

In exploring the therapeutic potential of these cytokines, it is important to note that while IL-17 inhibitors have shown promise, clinical trials have yielded mixed results. Factors contributing to these outcomes may include the complexity of the cytokine networks involved in RA, where IL-17 alone might not dominate the inflammatory landscape as it does in other diseases [66].

TARGETING ANTI-GRANULOCYTE MACROPHAGE-COLONY-STIMULATING FACTOR

GM-CSF is a cytokine that plays a pivotal role in the survival, proliferation, and activation of myeloid cells, including macrophages and neutrophils, which are key players in the inflammatory processes observed in rheumatoid arthritis (RA). GM-CSF enhances the production of other pro-inflammatory cytokines and chemokines, contributing to synovial inflammation, joint destruction, and systemic inflammation [60].

Several monoclonal antibodies targeting GM-CSF or its receptor (GM-CSFR) have been developed and tested in clinical trials for RA. Among these, mavrilimumab and otilimab are two prominent agents. Mavrilimumab is a human monoclonal antibody that

binds to the alpha subunit of the GM-CSF receptor, thereby preventing GM-CSF from activating its receptor on immune cells. Otilimab (GSK3196165) targets GM-CSF itself, neutralizing its activity and thereby reducing inflammation [61].

Clinical trials for these anti-GM-CSF therapies have shown promising results. Mavrilimumab has demonstrated significant reductions in disease activity and improvements in clinical outcomes compared to placebo in patients with moderate to severe RA. Patients treated with mavrilimumab showed decreased swollen and tender joint counts, reduced pain and disability scores, and lower levels of acute-phase reactants such as C-reactive protein (CRP). Similarly, otilimab has shown efficacy in reducing RA symptoms and improving physical function, with clinical trials indicating a favorable safety profile [62].

Therapeutic methods involving anti-GM-CSF agents typically include subcutaneous injections administered at regular intervals, such as every two to four weeks. The dosing regimen and duration of treatment can vary based on the specific drug, patient response, and disease severity. These therapies are generally considered for patients who have not responded adequately to traditional DMARDs or other biologic agents, such as TNF- α inhibitors [63].

LESSONS FROM FAILED CYTOKINE TARGETS IN RHEUMATOID ARTHRITIS

Another failed target is IL-20. This cytokine has been found to be elevated in patients with rheumatoid arthritis (RA), and preclinical studies suggested that it could contribute to joint inflammation and damage. However, clinical trials involving anti-IL-20 therapies did not demonstrate significant improvements in disease outcomes. Similarly, IL-21 was targeted due to its involvement in B cell differentiation and antibody production, yet anti-IL-21 therapies did not yield the expected clinical benefits. This may be due to

redundancy within cytokine networks or compensatory mechanisms that mitigate the effects of IL-21 inhibition [67].

IL-32 was also considered a promising target given its elevated levels in RA synovial tissue and its ability to induce other pro-inflammatory cytokines. However, clinical trials targeting IL-32 have not progressed far, likely due to insufficient efficacy or challenges in developing specific inhibitors. IL-33 and its receptor ST2 have also attracted attention because of their pro-inflammatory roles in RA. Despite preclinical data supporting their involvement in RA pathology, clinical attempts to block IL-33/ST2 signaling have not achieved significant success [68].

Granulocyte colony-stimulating factor (G-CSF) and macrophage migration inhibitory factor (MIF) were also explored as potential targets. G-CSF is involved in the production and activation of neutrophils, which contribute to RA inflammation. However, targeting G-CSF did not result in meaningful clinical improvements, possibly due to its essential role in normal immune function and the risk of impairing host defense. MIF, a cytokine that promotes inflammation and is elevated in RA, was another target that failed to translate into successful clinical outcomes, likely because of the complex interplay of cytokines in RA and compensatory mechanisms that limit the effects of MIF inhibition [69,70].

Inhibition of interleukin-15 (IL-15) was another approach that did not yield the desired results in RA treatment. Although IL-15 is involved in T cell activation and survival, clinical trials with IL-15 inhibitors did not demonstrate significant efficacy, highlighting the redundancy and adaptability of the immune system. Additionally, interleukin-22 (IL-22) was targeted due to its role in tissue remodeling and inflammation; however, clinical efforts to inhibit IL-22 did not produce significant therapeutic benefits in RA [71].

Table 2. Challenges and Lessons Learned from Cytokine Targeting in RA

Target Cytokine	Clinical Outcome	Reasons for Failure
IL-20	No significant improvement in disease outcomes	Lack of efficacy in clinical trials
IL-21	Did not yield expected clinical benefits	Redundancy within cytokine networks
IL-32	Limited progress in trials	Insufficient efficacy, challenges in inhibitor development
IL-33	Targeting failed to show success	Poor translation from preclinical to clinical success
G-CSF	Failed to improve clinical outcomes	Critical role in normal immune function
MIF	Did not translate into successful outcomes	Compensatory mechanisms limited efficacy
IL-15	Lack of significant efficacy in trials	Immune system adaptability dulled therapeutic effects
IL-22	Clinical attempts showed no significant benefits	Complex pathophysiological interplay

CONCLUSION

The exploration of cytokine-targeted therapies for rheumatoid arthritis (RA) presents a compelling approach to managing this complex and debilitating condition. Cytokines such as tumor necrosis factor- α (TNF- α), interleukin-6 (IL-6), interleukin-1 (IL-1), interleukin-17 (IL-17), interleukin-23 (IL-23), and granulocyte macrophage-colony stimulating factor (GM-CSF) have been the focus of extensive research and clinical investigation due to their pivotal roles in RA pathogenesis.

Biologic agents targeting these cytokines have demonstrated efficacy in reducing inflammation, alleviating disease activity, and improving clinical outcomes in many RA patients who do not adequately respond to traditional disease-modifying antirheumatic drugs (DMARDs). However, the variability of responses and the complex interactions within cytokine networks highlight the need for personalized treatment approaches and the exploration of additional therapeutic targets.

The lessons learned from failed cytokine targets underscore the challenges involved in translating preclinical promise into clinical success and emphasize the intricate nature of RA pathophysiology. Despite these setbacks, the pursuit of novel cytokine targets and a deeper understanding of their interactions offer opportunities to refine treatment strategies and enhance patient care.

In summary, the autoimmune origin of RA involves a multifaceted interplay between genetic factors, environmental triggers, and an aberrant immune response characterized by the dysregulation of key cytokines. Understanding the cross-talk between these cytokines and their role in perpetuating autoimmunity provides new insights into potential therapeutic strategies. The ongoing exploration of cytokine-targeted therapies underscores the exciting potential for personalized and effective treatment modalities in RA management.

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Writing—original draft preparation, A.V.P.; writing—review and editing, A.N.O, V.A.O., U.R., O.G., S.P. All authors have read and agreed to the published version of the manuscript.

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The authors declare no conflict of interest.

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REFERENCES

1. Chauhan K, Jandu JS, Brent LH, et al. Rheumatoid Arthritis. [Updated 2023 May 25]. In: StatPearls [Internet]. Treasure Island (FL): StatPearls Publishing; 2024 Jan-. Available from: <https://www.ncbi.nlm.nih.gov/books/NBK441999/>
2. Amaya-Amaya J, Rojas-Villarraga A, Mantilla RD, et al. Rheumatoid arthritis. In: Anaya JM, Shoenfeld Y, Rojas-Villarraga A, et al., editors. Autoimmunity: From Bench to Bedside [Internet]. Bogota (Colombia): El Rosario University Press; 2013 Jul 18. Chapter 24. Available from: <https://www.ncbi.nlm.nih.gov/books/NBK459454/>
3. Sweeney, S. E., & Firestein, G. S. (2004). Rheumatoid arthritis: regulation of synovial inflammation. *The international journal of biochemistry & cell biology*, 36(3), 372–378. [https://doi.org/10.1016/s1357-2725\(03\)00259-0](https://doi.org/10.1016/s1357-2725(03)00259-0)
4. Wang, Z., Wang, J., Lan, T., Zhang, L., Yan, Z., Zhang, N., Xu, Y., & Tao, Q. (2023). Role and mechanism of fibroblast-activated protein- α expression on the surface of fibroblast-like synoviocytes in rheumatoid arthritis. *Frontiers in immunology*, 14, 1135384. <https://doi.org/10.3389/fimmu.2023.1135384>
5. Atta, A., Salem, M. M., El-Said, K. S., & Mohamed, T. M. (2024). Mechanistic role of quercetin as inhibitor for adenosine deaminase enzyme in rheumatoid arthritis: systematic review. *Cellular & molecular biology letters*, 29(1), 14. <https://doi.org/10.1186/s11658-024-00531-7>
6. Tenazinha, C., Barros, R., Fonseca, J. E., & Vieira-Sousa, E. (2022). Histopathology of Psoriatic Arthritis Synovium-A Narrative Review. *Frontiers in medicine*, 9, 860813. <https://doi.org/10.3389/fmed.2022.860813>
7. Liu, S., Deng, Z., Chen, K., Jian, S., Zhou, F., Yang, Y., Fu, Z., Xie, H., Xiong, J., & Zhu, W. (2022). Cartilage tissue engineering: From proinflammatory and anti-inflammatory cytokines to osteoarthritis treatments (Review). *Molecular medicine reports*, 25(3), 99. <https://doi.org/10.3892/mmr.2022.12615>
8. Manosalva, C., Alarcon, P., Quiroga, J., Teuber, S., Carretta, M. D., Bustamante, H., Lopez-Muñoz, R., Hidalgo, M. A., & Burgos, R. A. (2023). Bovine tumor necrosis factor- α Increases IL-6, IL-8, and PGE2 in

- bovine fibroblast-like synoviocytes by metabolic reprogramming. *Scientific reports*, 13(1), 3257. <https://doi.org/10.1038/s41598-023-29851-y>
9. Panagopoulos, P. K., & Lambrou, G. I. (2018). Bone erosions in rheumatoid arthritis: recent developments in pathogenesis and therapeutic implications. *Journal of musculoskeletal & neuronal interactions*, 18(3), 304–319.
10. Kondo, N., Kuroda, T., & Kobayashi, D. (2021). Cytokine Networks in the Pathogenesis of Rheumatoid Arthritis. *International journal of molecular sciences*, 22(20), 10922. <https://doi.org/10.3390/ijms222010922>
11. Huang, J., Fu, X., Chen, X., Li, Z., Huang, Y., & Liang, C. (2021). Promising Therapeutic Targets for Treatment of Rheumatoid Arthritis. *Frontiers in immunology*, 12, 686155. <https://doi.org/10.3389/fimmu.2021.686155>
12. Lubberts E, van den Berg WB. Cytokines in the Pathogenesis of Rheumatoid Arthritis and Collagen-Induced Arthritis. In: Madame Curie Bioscience Database [Internet]. Austin (TX): Landes Bioscience; 2000-2013. Available from: <https://www.ncbi.nlm.nih.gov/books/NBK6288/>
13. Deckers, J., Anbergen, T., Hokke, A.M. *et al.* Engineering cytokine therapeutics. *Nat Rev Bioeng* 1, 286–303 (2023). <https://doi.org/10.1038/s44222-023-00030-y>
14. Jang, S., Kwon, E. J., & Lee, J. J. (2022). Rheumatoid Arthritis: Pathogenic Roles of Diverse Immune Cells. *International journal of molecular sciences*, 23(2), 905. <https://doi.org/10.3390/ijms23020905>
15. Zhang, C. (2021). Flare-up of cytokines in rheumatoid arthritis and their role in triggering depression: Shared common function and their possible applications in treatment (Review). *Biomedical Reports*, 14, 16. <https://doi.org/10.3892/br.2020.1392>
16. Ruiz de Morales, J. M. G., Puig, L., Daudén, E., Cañete, J. D., Pablos, J. L., Martín, A. O., Juanatey, C. G., Adán, A., Montalbán, X., Borruel, N., Ortí, G., Holgado-Martín, E., García-Vidal, C., Vizcaya-Morales, C., Martín-Vázquez, V., & González-Gay, M. Á. (2020). Critical role of interleukin (IL)-17 in inflammatory and immune disorders: An updated review of the evidence focusing in controversies. *Autoimmunity reviews*, 19(1), 102429. <https://doi.org/10.1016/j.autrev.2019.102429>
17. Torequl Islam, M., Quispe, C., Herrera-Bravo, J., Rahaman, M. M., Hossain, R., Sarkar, C., Raihan, M. A., Chowdhury, M. M., Uddin, S. J., Shilpi, J. A., Marcelo de Castro E Sousa, J., Melo-Cavalcante, A. A. C., Mubarak, M. S., Sharifi-Rad, J., & Calina, D. (2022). Activities and Molecular Mechanisms of Diterpenes, Diterpenoids, and Their Derivatives in Rheumatoid Arthritis. *Evidence-based complementary and alternative medicine : eCAM*, 2022, 4787643. <https://doi.org/10.1155/2022/4787643>
18. Deshpande, P., Cavanagh, M. M., Le Saux, S., Singh, K., Weyand, C. M., & Goronzy, J. J. (2013). IL-7- and IL-15-mediated TCR sensitization enables T cell responses to self-antigens. *Journal of immunology (Baltimore, Md. : 1950)*, 190(4), 1416–1423. <https://doi.org/10.4049/jimmunol.1201620>
19. Muth, K. N., Rech, J., Losch, F. O., & Hoerning, A. (2023). Reversing the Inflammatory Process-25 Years of Tumor Necrosis Factor- α Inhibitors. *Journal of clinical medicine*, 12(15), 5039. <https://doi.org/10.3390/jcm12155039>
20. Hong, K. H., Cho, M. L., Min, S. Y., Shin, Y. J., Yoo, S. A., Choi, J. J., Kim, W. U., Song, S. W., & Cho, C. S. (2007). Effect of interleukin-4 on vascular endothelial growth factor production in rheumatoid synovial fibroblasts. *Clinical and experimental immunology*, 147(3), 573–579. <https://doi.org/10.1111/j.1365-2249.2006.03295.x>
21. Szekanecz, Z., & Koch, A. E. (2009). Angiogenesis and its targeting in rheumatoid arthritis. *Vascular pharmacology*, 51(1), 1–7. <https://doi.org/10.1016/j.vph.2009.02.002>
22. Sarrand, J., & Soyfoo, M. S. (2023). Involvement of Epithelial-Mesenchymal Transition (EMT) in Autoimmune Diseases. *International journal of molecular sciences*, 24(19), 14481. <https://doi.org/10.3390/ijms241914481>
23. Alunno, A., Carubbi, F., Giacomelli, R. *et al.* Cytokines in the pathogenesis of rheumatoid arthritis: new players and therapeutic targets. *BMC Rheumatol* 1, 3 (2017). <https://doi.org/10.1186/s41927-017-0001-8>
24. Negrei, C., Bojinca, V., Balanescu, A., Bojinca, M., Baconi, D., Spandidos, D. A., Tsatsakis, A. M., & Stan, M. (2016). Management of rheumatoid arthritis: Impact and risks of various therapeutic approaches. *Experimental and therapeutic medicine*, 11(4), 1177–1183. <https://doi.org/10.3892/etm.2016.3045>
25. Ben Mrid, R., Bouchmaa, N., Ainani, H., El Fatimy, R., Malka, G., & Mazini, L. (2022). Anti-rheumatoid drugs advancements: New insights into the molecular treatment of rheumatoid arthritis. *Biomedicine & pharmacotherapy = Biomedecine & pharmacotherapie*, 151, 113126.

- <https://doi.org/10.1016/j.biopha.2022.113126>
26. Patel, A. M., & Moreland, L. W. (2010). Interleukin-6 inhibition for treatment of rheumatoid arthritis: a review of tocilizumab therapy. *Drug design, development and therapy*, 4, 263–278.
<https://doi.org/10.2147/DDDT.S14099>
27. Horton, S., Buch, M. H., & Emery, P. (2010). Efficacy, tolerability and safety of biologic therapy in rheumatoid disease: patient considerations. *Drug, healthcare and patient safety*, 2, 101–119.
<https://doi.org/10.2147/DHPS.S6317>
28. Senolt L. (2019). Emerging therapies in rheumatoid arthritis: focus on monoclonal antibodies. *F1000Research*, 8, F1000 Faculty Rev-1549.
<https://doi.org/10.12688/f1000research.18688.1>
29. Fagerli, K. M., Kearsley-Fleet, L., Watson, K. D., Packham, J., Contributors Group, B. R., Symmons, D. P. M., & Hyrich, K. L. (2018). Long-term persistence of TNF-inhibitor treatment in patients with psoriatic arthritis. Data from the British Society for Rheumatology Biologics Register. *RMD open*, 4(1), e000596.
<https://doi.org/10.1136/rmdopen-2017-000596>
30. Escudero-Vilaplana, V., Ramírez-Herraiz, E., Trovato-López, N., Alañón-Plaza, E., Bellini, M. J., Herranz-Alonso, A., Bellón-Cano, J. M., Morell-Baladrón, A., & Sanjurjo-Sáez, M. (2012). Influence on effectiveness of early treatment with anti-TNF therapy in rheumatoid arthritis. *Journal of pharmacy & pharmaceutical sciences : a publication of the Canadian Society for Pharmaceutical Sciences, Societe canadienne des sciences pharmaceutiques*, 15(3), 355–360.
<https://doi.org/10.18433/j33w30>
31. Zhang, H., Shi, N., Diao, Z., Chen, Y., & Zhang, Y. (2020). Therapeutic potential of TNF α inhibitors in chronic inflammatory disorders: Past and future. *Genes & diseases*, 8(1), 38–47.
<https://doi.org/10.1016/j.gendis.2020.02.004>
32. Goel, N., & Stephens, S. (2010). Certolizumab pegol. *mAbs*, 2(2), 137–147.
<https://doi.org/10.4161/mabs.2.2.11271>
33. Li, L., Zhang, Y., Ma, L., Ji, P., Yim, S., Chowdhury, B., Doddapaneni, S., Liu, J., Wang, Y., & Sahajwalla, C. (2017). Exposure-Response Modeling and Power Analysis of Components of ACR Response Criteria in Rheumatoid Arthritis (Part 1: Binary Model). *Journal of clinical pharmacology*, 57(9), 1097–1106.
<https://doi.org/10.1002/jcph.891>
34. Ward, M. M., Guthrie, L. C., & Alba, M. I. (2014). Brief report: rheumatoid arthritis response criteria and patient-reported improvement in arthritis activity: is an American College of Rheumatology twenty percent response meaningful to patients?. *Arthritis & rheumatology (Hoboken, N.J.)*, 66(9), 2339–2343.
<https://doi.org/10.1002/art.38705>
35. Takeuchi, T., Kawanishi, M., Nakanishi, M., Yamasaki, H., & Tanaka, Y. (2022). Phase II/III Results of a Trial of Anti-Tumor Necrosis Factor Multivalent NANOBODY Compound Ozoralizumab in Patients With Rheumatoid Arthritis. *Arthritis & rheumatology (Hoboken, N.J.)*, 74(11), 1776–1785.
<https://doi.org/10.1002/art.42273>
36. Donahue, K. E., Schulman, E. R., Gartlehner, G., Jonas, B. L., Coker-Schwimmer, E., Patel, S. V., Weber, R. P., Bann, C. M., & Viswanathan, M. (2019). Comparative Effectiveness of Combining MTX with Biologic Drug Therapy Versus Either MTX or Biologics Alone for Early Rheumatoid Arthritis in Adults: a Systematic Review and Network Meta-analysis. *Journal of general internal medicine*, 34(10), 2232–2245.
<https://doi.org/10.1007/s11606-019-05230-0>
37. Menegatti, S., Bianchi, E., & Rogge, L. (2019). Anti-TNF Therapy in Spondyloarthritis and Related Diseases, Impact on the Immune System and Prediction of Treatment Responses. *Frontiers in immunology*, 10, 382.
<https://doi.org/10.3389/fimmu.2019.00382>
38. Venkatesha, S. H., Dudics, S., Acharya, B., & Moudgil, K. D. (2014). Cytokine-modulating strategies and newer cytokine targets for arthritis therapy. *International journal of molecular sciences*, 16(1), 887–906.
<https://doi.org/10.3390/ijms16010887>
39. Soler, M. F., Abaurrea, A., Azcoaga, P., Araujo, A. M., & Caffarel, M. M. (2023). New perspectives in cancer immunotherapy: targeting IL-6 cytokine family. *Journal for immunotherapy of cancer*, 11(11), e007530.
<https://doi.org/10.1136/jitc-2023-007530>
40. Miao, P., Zhou, X. W., Wang, P., Zhao, R., Chen, N., Hu, C. Y., Chen, X. H., Qian, L., Yu, Q. W., Zhang, J. Y., Xu, R., He, D. Y., Xiao, L. B., Li, P., Lu, M., & Zhang, D. Q. (2018). Regulatory effect of anti-gp130 functional mAb on IL-6 mediated RANKL and Wnt5a expression through JAK-STAT3 signaling pathway in FLS. *Oncotarget*, 9(29), 20366–20376.
<https://doi.org/10.18632/oncotarget.23917>
41. Favalli E. G. (2020). Understanding the Role of Interleukin-6 (IL-6) in the Joint and Beyond: A Comprehensive Review of IL-6 Inhibition for the Management of Rheumatoid

- Arthritis. *Rheumatology and therapy*, 7(3), 473–516. <https://doi.org/10.1007/s40744-020-00219-2>
42. Mihara, M., Ohsugi, Y., & Kishimoto, T. (2011). Tocilizumab, a humanized anti-interleukin-6 receptor antibody, for treatment of rheumatoid arthritis. *Open access rheumatology : research and reviews*, 3, 19–29. <https://doi.org/10.2147/OARRR.S17118>
43. Burmester, G. R., Lin, Y., Patel, R., van Adelsberg, J., Mangan, E. K., Graham, N. M., van Hoogstraten, H., Bauer, D., Ignacio Vargas, J., & Lee, E. B. (2017). Efficacy and safety of sarilumab monotherapy versus adalimumab monotherapy for the treatment of patients with active rheumatoid arthritis (MONARCH): a randomised, double-blind, parallel-group phase III trial. *Annals of the rheumatic diseases*, 76(5), 840–847. <https://doi.org/10.1136/annrheumdis-2016-210310>
44. Krumm, B., Xiang, Y., & Deng, J. (2014). Structural biology of the IL-1 superfamily: key cytokines in the regulation of immune and inflammatory responses. *Protein science : a publication of the Protein Society*, 23(5), 526–538. <https://doi.org/10.1002/pro.2441>
45. Goldbach-Mansky R. (2009). Blocking interleukin-1 in rheumatic diseases. *Annals of the New York Academy of Sciences*, 1182, 111–123. <https://doi.org/10.1111/j.1749-6632.2009.05159.x>
46. Goh, A. X., Bertin-Maghit, S., Ping Yeo, S., Ho, A. W., Derks, H., Mortellaro, A., & Wang, C. I. (2014). A novel human anti-interleukin-1 β neutralizing monoclonal antibody showing in vivo efficacy. *mAbs*, 6(3), 765–773. <https://doi.org/10.4161/mabs.28614>
47. Cavalli, G., & Dinarello, C. A. (2015). Treating rheumatological diseases and comorbidities with interleukin-1 blocking therapies. *Rheumatology (Oxford, England)*, 54(12), 2134–2144. <https://doi.org/10.1093/rheumatology/kev269>
48. Jarlborg, M., & Gabay, C. (2022). Systemic effects of IL-6 blockade in rheumatoid arthritis beyond the joints. *Cytokine*, 149, 155742. <https://doi.org/10.1016/j.cyto.2021.155742>
49. Wang, R., & Maksymowych, W. P. (2021). Targeting the Interleukin-23/Interleukin-17 Inflammatory Pathway: Successes and Failures in the Treatment of Axial Spondyloarthritis. *Frontiers in immunology*, 12, 715510. <https://doi.org/10.3389/fimmu.2021.715510>
50. Meher, J., Patel, S., Nanda, R., & Siddiqui, M. S. (2023). Association of Serum IL-17 and IL-23 Cytokines With Disease Activity and Various Parameters of Rheumatoid Arthritis in Indian Patients. *Cureus*, 15(11), e49654. <https://doi.org/10.7759/cureus.49654>
51. Molnar, V., Matišić, V., Kodvanj, I., Bjelica, R., Jeleč, Ž., Hudetz, D., Rod, E., Čukelj, F., Vrdoljak, T., Vidović, D., Starešinić, M., Sabalić, S., Dobričić, B., Petrović, T., Antičević, D., Borić, I., Košir, R., Zmrzljak, U. P., & Primorac, D. (2021). Cytokines and Chemokines Involved in Osteoarthritis Pathogenesis. *International journal of molecular sciences*, 22(17), 9208. <https://doi.org/10.3390/ijms22179208>
52. Silvagni, E., Missiroli, S., Perrone, M., Patergnani, S., Boncompagni, C., Bortoluzzi, A., Govoni, M., Giorgi, C., Alivernini, S., Pinton, P., & Scirè, C. A. (2021). From Bed to Bench and Back: TNF- α , IL-23/IL-17A, and JAK-Dependent Inflammation in the Pathogenesis of Psoriatic Synovitis. *Frontiers in pharmacology*, 12, 672515. <https://doi.org/10.3389/fphar.2021.672515>
53. Țiburcă, L., Bembea, M., Zaha, D. C., Jurca, A. D., Vesa, C. M., Rațiu, I. A., & Jurca, C. M. (2022). The Treatment with Interleukin 17 Inhibitors and Immune-Mediated Inflammatory Diseases. *Current issues in molecular biology*, 44(5), 1851–1866. <https://doi.org/10.3390/cimb44050127>
54. Sánchez-Rodríguez, G., & Puig, L. (2023). Pathogenic Role of IL-17 and Therapeutic Targeting of IL-17F in Psoriatic Arthritis and Spondyloarthropathies. *International journal of molecular sciences*, 24(12), 10305. <https://doi.org/10.3390/ijms241210305>
55. McGonagle, D. G., McInnes, I. B., Kirkham, B. W., Sherlock, J., & Moots, R. (2019). The role of IL-17A in axial spondyloarthritis and psoriatic arthritis: recent advances and controversies. *Annals of the rheumatic diseases*, 78(9), 1167–1178. <https://doi.org/10.1136/annrheumdis-2019-215356>
56. Globig, A. M., Sommer, N. P., Wild, K., Schardey, J., Zoldan, K., Thomann, A. K., Schulte, L. A., Schreiner, R., Reindl, W., Klaus, J., Schempp, C. M., Hofmann, M., Thimme, R., Boettler, T., & Hasselblatt, P. (2021). Ustekinumab Inhibits T Follicular Helper Cell Differentiation in Patients With Crohn's Disease. *Cellular and molecular gastroenterology and hepatology*, 11(1), 1–12. <https://doi.org/10.1016/j.jcmgh.2020.07.005>
57. Kavanaugh, A., Ritchlin, C., Rahman, P., Puig, L., Gottlieb, A. B., Li, S., Wang, Y., Noonan, L., Brodmerkel, C., Song, M., Mendelsohn, A. M., McInnes, I. B., & PSUMMIT-1 and 2 Study Groups (2014). Ustekinumab, an anti-IL-12/23 p40 monoclonal antibody, inhibits radiographic progression in patients with active psoriatic arthritis: results of an integrated analysis of

- radiographic data from the phase 3, multicentre, randomised, double-blind, placebo-controlled PSUMMIT-1 and PSUMMIT-2 trials. *Annals of the rheumatic diseases*, 73(6), 1000–1006. <https://doi.org/10.1136/annrheumdis-2013-204741>
58. Sawyer, L. M., Malottki, K., Sabry-Grant, C., Yasmeen, N., Wright, E., Sohr, A., Borg, E., & Warren, R. B. (2019). Assessing the relative efficacy of interleukin-17 and interleukin-23 targeted treatments for moderate-to-severe plaque psoriasis: A systematic review and network meta-analysis of PASI response. *PloS one*, 14(8), e0220868. <https://doi.org/10.1371/journal.pone.0220868>
59. Tsai, Y. C., & Tsai, T. F. (2017). Anti-interleukin and interleukin therapies for psoriasis: current evidence and clinical usefulness. *Therapeutic advances in musculoskeletal disease*, 9(11), 277–294. <https://doi.org/10.1177/1759720X17735756>
60. Lotfi, N., Thome, R., Rezaei, N., Zhang, G. X., Rezaei, A., Rostami, A., & Esmail, N. (2019). Roles of GM-CSF in the Pathogenesis of Autoimmune Diseases: An Update. *Frontiers in immunology*, 10, 1265. <https://doi.org/10.3389/fimmu.2019.01265>
61. Burmester, G. R., Feist, E., Sleeman, M. A., Wang, B., White, B., & Magrini, F. (2011). Mavrilimumab, a human monoclonal antibody targeting GM-CSF receptor- α , in subjects with rheumatoid arthritis: a randomised, double-blind, placebo-controlled, phase I, first-in-human study. *Annals of the rheumatic diseases*, 70(9), 1542–1549. <https://doi.org/10.1136/ard.2010.146225>
62. Taylor, P. C., Weinblatt, M. E., McInnes, I. B., Atsumi, T., Strand, V., Takeuchi, T., Bracher, M., Brooks, D., Davies, J., Goode, C., Gupta, A., Mukherjee, S., O'Shea, C., Saurigny, D., Schifano, L. A., Shelton, C., Smith, J. E., Wang, M., Wang, R., Watts, S., ... Fleischmann, R. M. (2023). Anti-GM-CSF otilimab versus sarilumab or placebo in patients with rheumatoid arthritis and inadequate response to targeted therapies: a phase III randomised trial (contRAst 3). *Annals of the rheumatic diseases*, 82(12), 1527–1537. <https://doi.org/10.1136/ard-2023-224449>
63. Zhang, F., Weng, D., Su, Y., Yin, C., Shen, L., Zhang, Y., Zhou, Y., Li, Q., Hu, Y., & Li, H. (2020). Therapeutic effect of subcutaneous injection of low dose recombinant human granulocyte-macrophage colony-stimulating factor on pulmonary alveolar proteinosis. *Respiratory research*, 21(1), 1. <https://doi.org/10.1186/s12931-019-1261-1>
64. Zhu, M., Ding, Q., Lin, Z., Fu, R., Zhang, F., Li, Z., Zhang, M., & Zhu, Y. (2023). New Targets and Strategies for Rheumatoid Arthritis: From Signal Transduction to Epigenetic Aspect. *Biomolecules*, 13(5), 766. <https://doi.org/10.3390/biom13050766>
65. Zimmerman, D. H., Szekanecz, Z., Markovics, A., Rosenthal, K. S., Carambula, R. E., & Mikecz, K. (2024). Current status of immunological therapies for rheumatoid arthritis with a focus on antigen-specific therapeutic vaccines. *Frontiers in immunology*, 15, 1334281. <https://doi.org/10.3389/fimmu.2024.1334281>
66. McGeachy, M. J., Cua, D. J., & Gaffen, S. L. (2019). The IL-17 Family of Cytokines in Health and Disease. *Immunity*, 50(4), 892–906. <https://doi.org/10.1016/j.immuni.2019.03.021>
67. Kragstrup, T. W., Andersen, T., Heftdal, L. D., Hvid, M., Gerwien, J., Sivakumar, P., Taylor, P. C., Senolt, L., & Deleuran, B. (2018). The IL-20 Cytokine Family in Rheumatoid Arthritis and Spondyloarthritis. *Frontiers in immunology*, 9, 2226. <https://doi.org/10.3389/fimmu.2018.02226>
68. Kwon, O. C., Park, M. C., & Kim, Y. G. (2023). Interleukin-32 as a biomarker in rheumatic diseases: A narrative review. *Frontiers in immunology*, 14, 1140373. <https://doi.org/10.3389/fimmu.2023.1140373>
69. Christensen, A. D., Haase, C., Cook, A. D., & Hamilton, J. A. (2016). K/BxN Serum-Transfer Arthritis as a Model for Human Inflammatory Arthritis. *Frontiers in immunology*, 7, 213. <https://doi.org/10.3389/fimmu.2016.00213>
70. Schindler, L., Smyth, L. C. D., Bernhagen, J., Hampton, M. B., & Dickerhof, N. (2021). Macrophage migration inhibitory factor (MIF) enhances hypochlorous acid production in phagocytic neutrophils. *Redox biology*, 41, 101946. <https://doi.org/10.1016/j.redox.2021.101946>
71. Guo, Y., Luan, L., Patil, N. K., & Sherwood, E. R. (2017). Immunobiology of the IL-15/IL-15R α complex as an antitumor and antiviral agent. *Cytokine & growth factor reviews*, 38, 10–21. <https://doi.org/10.1016/j.cytogfr.2017.08.002>